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(54) Title: CANDIDA ALBICANS PROTEINS ASSOCIATED WITH VIRULENCE AND HYPHAL FORMATION AND USES THEREOF

(57) Abstract

The present invention relates to Candida albicans proteins, such as CaCla4p, Cst20p, CaCdc42p and CaBemlp, associated with virulence and hyphal formation and uses thereof, such as to design screening tests for inhibitors for the treatment of pathogenic fungi infections and/or inflammation conditions. The invention also relates to an in vitro screening test for compounds to inhibit the biological activity of at least one protein selected from the group consisting of CaCla4p, Cst20p, CaCdc42p and CaBemlp, which comprises: a) at least one of said proteins; and b) means to monitor the biological activity of said at least one protein; thereby compounds are tested for their inhibiting potential.

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CANDIDA ALBICANS PROTEINS ASSOCIATED WITH VIRULENCE AND HYPHAL FORMATION AND USES THEREOF

BACKGROUND OF THE INVENTION

(a) Field of the Invention

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The invention relates to Candida albicans proteins, such as CaCla4p, Cst20p, CaCdc42p and CaBemlp, associated with virulence and hyphal formation and uses thereof, such as to design screening tests for inhibitors for the treatment of pathogenic fungi infections and/or inflammation conditions.

(b) Description of Prior Art

Candida albicans is the major fungal pathogen in humans, causing various forms of candidiasis. incidence of infections is increasing in immunocompromised patients. This fungus is diploid with no sexual cycle and is capable of a morphological transition from a unicellular budding yeast to a filamentous form. Extensive filamentous growth leads to the formation of a mycelium displaying hyphae with branches and lateral buds. In view of the observation that hyphae seem to adhere to and invade host tissues more readily than does the yeast form, the switch from the yeast to the filamentous form probably contributes to the virulence of this organism (for a review see Fidel, P. L. & Sobel, J. D. (1994) Trends Microbiol. 2, 202-205). The molecular mechanisms by which morphological switching is regulated are poorly understood.

Like C. albicans, bakers yeast Saccharomyces

30 cerevisiae is also a dimorphic organism capable of switching under certain nutritional conditions from a budding yeast to a filamentous form. Under the control of nutritional signals, diploid cells switch to pseudohyphal growth (Gimeno, C. J. et al. (1992) Cell 68,

35 1077-1090), and haploid cells to invasive growth

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(Roberts, R. L. & Fink, G. R. (1994) Genes Dev. 8, 2974-2985).

The similarities between the dimorphic switching of S. cerevisiae and C. albicans suggest that these 5 morphological pathways may be regulated by similar mechanisms in both organisms. In S. cerevisiae, morphological transitions are controlled by signaling components that are also involved in the mating response of haploid cells (Roberts, R. L. & Fink, G. R. (1994) 10 Genes Dev. 8, 2974-2985; Liu, H. et al. (1993) Science 262, 1741-1744). The switch to pseudohyphal growth requires a transcription factor encoded by the STE12 gene, and a mitogen-activated protein (MAP) kinase cascade including Ste7p (a homolog of MAP kinase kinase or MEK), Stellp (a MEK kinase homolog) and Ste20p (a MEK 15 kinase kinase) (Roberts, R. L. & Fink, G. R. (1994) Genes Dev. 8, 2974-2985; Liu, H. et al. (1993) Science The MAP kinases involved in this **262.** 1741-1744). response are as yet unknown (Roberts, R. L. & Fink, G. R. (1994) Genes Dev. 8, 2974-2985; Liu, H. et al. 20 (1993) Science 262, 1741-1744).

Members of the Ste20p family of serine/threonine protein kinases are thought to be involved in triggering morphogenetic processes in response to external signals in organisms ranging from yeast to mammalian cells. Two of these kinases, Ste20p and Cla4p, are well characterized in S. cerevisiae (Leberer, E. et al. (1992) EMBO J. 11, 4815-4824; Cvrckova, F. (1995) Genes Dev. 9, 1817-1830). Ste20p is required for pheromone signal transduction (Leberer, Ε. et (1992) EMBO J. 11, 4815-4824) and for filamentous growth in response to nitrogen starvation (Roberts, R. L. & Fink, G. R. (1994) Genes Dev. 8, 2974-2985; Liu, H. et al. (1993) Science 262, 1741-1744), and shares an essential function with Cla4p during budding (Cvrckova,

F. et al. (1995) Genes Dev. 9, 1817-1830). Ste20p and Cla4p interact with the small G-protein Cdc42p, and this interaction is required for viability of S. cerevisiae cells. Ste20p also interacts with the SH3 domain protein Bemlp, and this interaction plays a role in morphogenetic processes (Leeuw, T. et al. (1995) Science 270, 1210-1213).

Here we show that Cst20p, a *C. albicans* homolog of the Ste20p protein kinase, is required for hyphal growth of *C. albicans* under certain *in vitro* conditions. We also show in a mouse model for systemic candidiasis that Cst20p plays a role in virulence, as judged from significantly prolonged survival of mice infected with *CST20* deleted cells. Our results suggest that Cst20p acts in a regulatory pathway which is involved in hyphal growth of *C. albicans*.

We also demonstrate that CaCla4p, a *C. albicans* homolog of the Cla4p protein kinase, is required for hyphal formation *in vitro* in response to serum, and *in vivo* in a mouse model for systemic candidiasis. We also show that CaCla4p is required for efficient colonization of kidneys with *C. albicans* cells after infection of mice and essential for virulence in the mouse model.

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SUMMARY OF THE INVENTION

One aim of the present invention is to provide Candida albicans proteins, such as CaCla4p, Cst20p, CaCdc42p and CaBemlp, and their uses thereof.

One aim of the present invention is to provide the nucleotide and amino acid sequences of CaCla4p, Cst20p, CaCdc42p and CaBemlp.

Another aim of the present invention is to provide screening tests for inhibitors of CaCla4p, Cst20p, CaCdc42p and CaBemlp or of their interactions.

The term "fungi" when used herein is intended to mean any fungi, pathogenic or not, which show hyphal induction using kinases, such as C. albicans, Saccharomyces cerevisiae, Aspergillus, Ustilago maydis, and all the species of the fungal genera Aspergillus, Blastomyces, Candida, Cladosporium, Coccidioides, Cryptococcus, Epidermophyton, Exophilia, Fonsecaea, Histoplasma, Madurella, Malassezia, Microsporum, Paracoccidioides, Penicillium, Phaeoannellomyces, Phialophora, Scedosporium, Sporothrix, Torulopsis, Trichophyton, Trichosporon, Ustilago, Wangiella, Xylohypha, among others.

In accordance with the present invention there is provided an *in vitro* screening test for compounds to inhibit the biological activity of at least one protein selected from the group consisting of CaCla4p, Cst20p, Cdc42p and Bemlp, which comprises:

a) at least one of the proteins; and

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- b) means to monitor the biological activity of at least one protein;
- 20 thereby compounds are tested for their inhibiting potential.

In accordance with another embodiment of the present invention, the inhibition of the interactions between CaCla4p and CaCdc42p is determined.

In accordance with another embodiment of the present invention, the inhibition of the interactions between Cst20p and CaCdc42p is determined.

BRIEF DESCRIPTION OF THE DRAWINGS

Figs. 1A to 1D illustrate photomicrographs which show that *C. albicans CST20* gene complements defects in pseudohyphal growth of ste20/ste20 S. cerevisiae diploid cells.

Figs. 2A to 2C show the morphology of *S. cere-*35 *visiae MAT* α cells (strain YEL306-1A) deleted for *STE20*and *CLA4*, and transformed with plasmids expressing *CLA4*

(Fig. 2A), STE20 (Fig. 2B) and C. albicans CST20 (Fig. 2C).

Figs. 3A to 3C show the nucleotide (SEQ ID NO:5) and predicted amino acid sequences of CST20 (SEQ ID NO:6).

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Fig. 4A is the deletion of CST20 in C. albicans.

Fig. 4B is the Southern blot analysis with a

CST20 fragment from EcoRI to XbaI as a probe.

Figs. 5A to 5J show colonies of C. albicans 10 cells grown for 5 days at 37°C on solid "Spider" medium containing mannitol. Wild type strain SC5314 ura3/ura3 cst20\(\Delta/cst20\Delta::URA3\) strain CDH22 (B), ura3/ura3 cst20∆/cst20∆::CST20::URA3 strain CDH36 (obtained by reintegration of CST20 into strain CDH25 15 by homologous recombination using linearized plasmid pDH190) (C), ura3/ura3 cst20\(D)/cst20\(D) strain CDH25 transformed with plasmids pYPB1-ADHpt (D) and pYPB1-ADHpt-HST7 (E), ura3/ura3 hst7\(\Delta\)/hst7\(\Delta\) strain CDH12 transformed with plasmids pVEC (F), pVEC-HST7 (G), 20 pYPBl-ADHpt (H), and pyPBl-ADHpt-HST7 (I), ura3/ura3 cph1/cph1 strain CDH72 [ura3/ura3 derivative of strain JK19] transformed with pYPB1-ADHpt-HST7 (J). Photomicrographs of representative colonies were taken with a 2x lens (bar=2mm).

Figs. 6A to 6C illustrate virulence assays. Survival curves of mice (n=10 for each *C. albicans* strain at each inoculation dose) infected with 1 x 10⁶ (A) and 1 x 10⁵ (B) cells of *C. albicans* strains SC5314 (wild type), CAI4 (ura3/ura3), CDH22 (ura3/ura3 cst20Δ/cst20Δ ::URA3) (C) Staining of mouse kidney sections with periodic acid Schiff's stain 48 hours after infection with cst20Δ/cst20Δ::URA3 mutant strain CDH22 (a). Some hyphal cells are indicated with arrows (bar=0.1 mm).

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Figs. 7A to 7B illustrate the nucleotide (SEQ ID NO:7) and predicted amino acid (SEQ ID NO:8) sequences of CaCLA4.

Fig. 8A illustrates the deletion of CaCLA4 in C. 5 albicans.

Fig. 8B illustrates the Southern blot analysis with the CaCLA4 fragment from PstI to XbaI as a probe.

Fig. 8C illustrates the Northern blot analysis with the CaCLA4 fragment as a probe. PCR with the divergent oligodeoxynucleotides OEL109 and OEL110 was used to delete the coding sequence of CaCLA4. A hisG-URA3-hisG cassette was then inserted, and homologous recombination was used in a two-step procedure to replace both CaCLA4 alleles.

Fig. 9 illustrates virulence assays. Survival curves of mice (n=15 for each *C. albicans* strain) infected with 1 x 10⁶ cells of *C. albicans* strains SC5314 (wild-type), CDH77 (CaCLA4/cacla4Δ), CLJ1 (cacla4Δ/cacla4Δ) and CLJ5 (CaCla4Δ/cacla4Δ) transformed with the control plasmid pVEC and plasmid pVEC-CaCLA4 carrying the CaCLA4 gene.

Fig. 10 illustrates the staining of mouse kidney sections with periodic acid Schiff's stain 48 h after infection with *C. albicans* strains SC5314 and CLJ1.

Fig. 11 illustrates the nucleotide (SEQ ID NO:9) and predicted amino acid (SEQ ID NO:10) sequences of CaCdc42p.

Figs. 12A to 12B illustrate the nucleotide (SEQ ID NO:11) and predicted amino acid (SEQ ID NO:12) sequences of CaBemlp.

DETAILED DESCRIPTION OF THE INVENTION

The CST20 gene of Candida albicans was cloned by functional complementation of a deletion of the STE20 gene in Saccharomyces cerevisiae. CST20 encodes a homolog of the $Ste20p/p65^{PAK}$ family of protein kinases.

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of C. albicans cells deleted for CST20 revealed defects in the lateral formation of mycelia on synthetic solid "Spider" media. However, hyphal development was not impaired in some other media. deleted for CST20 were less virulent in a mouse model for systemic candidiasis. Our results suggest that more than one signaling pathway can trigger hyphal development in C. albicans, one of which has a protein kinase cascade that is analogous to the mating response pathway in S. cerevisiae and might have become adapted to the control of mycelial formation in C. albicans.

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The CaCLA4 gene of C. albicans was cloned by functional complementation of the growth defect of S. cerevisiae cells deleted for the STE20 and CLA4 genes. CaCLA4 encodes a homolog of the Ste20p family of serine/threonine protein kinases with pleckstrin homology and Cdc42p binding domains in the amino-terminal noncatalytic region. Deletion of both alleles of CaCLA4 in C. albicans caused defects in hyphal formation in vitro in synthetic liquid and solid media, and in vivo in a mouse model for systemic candidiasis. The deletions reduced the invasion of C. albicans cells into kidneys after infection into mice and completely suppressed virulence in the mouse model. Thus, hyphal formation of C. albicans mediated by the CaCla4p protein kinase may contribute to the pathogenicity of this dimorphic fungus.

The CaBEM1 and CaCDC42 genes of C. albicans were cloned by functional complementation of the growth defect of S. cerevisiae cells deleted for the BEM1 and CDC42 genes, respectively. CaBEM1 encodes an SH3 domain protein with homology to Bemlp, and CaCDC42 encodes a small G-protein with homology to members of the Rho-family of G-proteins.

MATERIALS AND METHODS

Yeast manipulations

The yeast form of C. albicans was cultured at 30°C in YPD medium. Hyphal growth was induced at 37°C 5 on solid "Spider" media (Liu, H. et al. (1994) Science 266, 1723-1726) containing 1% (w/v) nutrient broth, 0.2% (w/v) K_2HPO_4 , 2% (w/v) agar and 1% (w/v) of the indicated sugars (pH 7.2 after autoclaving). 10 were grown in liquid "Spider" media at 30°C to stationary phase, and then incubated for 5 days at 37°C on solid "Spider" media at a density of about 200 cells per 80 mm plates. All media were supplemented with uridine (25 μg/ml) for the growth of Ura strains. 15 Germ tube formation was induced at 37°C in either 10% fetal bovine serum (GIBCO/BRL) on liquid "Spider" media containing the indicated sugars at an inoculation density of 10⁷ cells per ml.

Yeast manipulations were performed according to 20 standard procedures.

Isolation of CST20

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The CST20 gene was isolated from a genomic C. albicans library constructed in plasmid YEp352 from genomic DNA of the clinical isolate WO1 (Boone, C. et al. (1991) J. Bacteriol. 173, 6859-6864). A plasmid carrying an amino-terminally truncated version of CST20 missing the first 918 nucleotides of coding sequence was isolated by screening for suppressors of defects in basal FUS1::HIS3 expression and mating in S. cerevisiae strain YEL64 which was disrupted in STE20. A fragment from nucleotides 958 to 1,252 of CST20 was amplified by the polymerase chain reaction (PCR) and used as a probe to isolate a full length clone by colony hybridization to the C. albicans genomic library transformed into E. coli strain MC1061. Both DNA strands were sequenced by

the dideoxy chain termination method. The full length clone was subcloned between the SacI and HindIII sites of the S. cerevisiae centromere plasmid pRS316 to yield plasmid pRL53.

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5 Isolation of CaCLA4

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The S. cerevisiae MATa strain YEL257-1A-2 deleted for STE20 and CLA4 and carrying plasmid pDH129 with CLA4 under control of the GAL1 promoter was transformed with the genomic C. albicans library constructed in the S. cerevisiae vector YEp352 carrying URA3 as selectable marker (Boone, C. et al. (1991) J. Bacte-173, 6859-6864). Transformants were grown on selective medium in 4% galactose and then replicaplated to selective medium containing 2% glucose to select for plasmids that were able to support growth in the absence of Cla4p and Ste20p. By screening 1,600 transformants, we isolated plasmid YEp352-CaCLA4 carrying an insert of 5.6 kb with an open reading frame of 2,913 bp capable of encoding a homolog of Cla4p. Subcloning indicated that this open reading frame was responsible for complementation. Both DNA strands were sequenced by the dideoxy chain termination method.

Molecular cloning of CaCDC42

the S. cerevisiae MATa strain DJTD2-16A carrying
the cdc42-1ts mutation was transformed with the genomic
C. albicans library constructed in the S. cerevisiae
vector YEp352 carrying URA3 as selectable marker
(Boone, C. et al. (1991) J. Bacteriol. 173, 6859-6864).
Transformants were grown on selective medium at room
temperature. Colonies were then replica-plated to
selective medium and grown at 34°C. By screening 2,000
transformants, we isolated plasmid YEp352-CaCDC42 carrying an open reading frame of 573 bp capable of encoding a homolog of Cdc42p. Both DNA strands were
sequenced by the dideoxy chain termination method. Sub-

cloning of various restriction endonuclease fragments indicated that the open reading frame was responsible for complementation of the temperature-sensitive growth defect caused by the cdc42-1^{ts} mutation.

5 Molecular cloning of CaBEM1

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The S. cerevisiae MATa strain YEL220-lA deleted for BEM1 and carrying plasmid pGAL-BEM1 with BEM1 under control of the GAL1 promoter was transformed with the genomic C. albicans library constructed in the S. cerevisiae vector YEp352 carrying URA3 as selectable marker (Boone, C. et al. (1991) J. Bacteriol. 173, 6859-6864). Transformants were grown on selective medium in 4% galactose and then replica-plated to selective medium containing 2% glucose to select for plasmids that were capable of supporting growth of Bemlp-depleted cells. We isolated plasmid YEp352-CaBEM1 carrying an open reading frame of 1,905 bp fulfilling this criterion and capable of encoding a homolog of Bemlp. Both DNA strands were sequenced by the dideoxy chain termination method, and subcloning of various restriction endonuclease fragments indicated that this open reading frame was responsible for complementation.

Construction of C. albicans strains and plasmids

To construct a CST20 null mutant, an EcoRI to SacI fragment from nucleotide positions 989 to 4,134 of CST20 was subcloned into the Bluescript KS(+) vector (Stratagene) to yield plasmid pDH119. A plasmid that contained CST20-flanking sequences from nucleotides 989 to 1,674, and 3,423 to 4,134 joined with BamHI sites, then created by PCR using the divergent oligodeoxynucleotide ODH68 (5'primers CGGGATCCAGACCAACCACTCGAACTACT-3' (SEO NO:1) ID and (5'-CGGGATCCGAAGGTGAACCACCATATTTG-3' ODH69 NO:2); newly introduced BamHI sites are underlined) and plasmid pDH119 as a template. The amplified DNA was

cleaved with BamHI and ligated with a 4 kb BamHI to BglII fragment of a hisG-URA3-hisG cassette derived from plasmid pCUB-6 (Fonzi, W. A. & Irwin, M. Y. (1993) Genetics 134, 717-728) to yield plasmid pDH183. plasmid was linearized with XhoI and SacI and transformed into the Ura C. albicans strain CAI4 (Fonzi, W. A. & Irwin, M. Y. (1993) Genetics 134, 717-728) to partially replace the coding region of one of the chromosomal CST20 alleles with the hisG-URA3-hisG cassette by Ura⁺ transformants homologous recombination. selected on Ura medium, and integration of the cassette into the CST20 locus was verified by Southern blot analysis. Spontaneous Ura derivatives of two of the heterozygous disruptants were selected on medium containing 5-fluoroorotic acid. These clones were screened by Southern blot hybridization to identify those which had lost the URA3 gene by intrachromosomal recombination mediated by the hisG repeats. This procedure was then repeated to delete the remaining functional allele of CST20.

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A similar procedure was employed to delete the CaCST20 gene. A 4.6 kb XbaI fragment of YEp352-CaCLA4 subcloned into the pBluescript KS(+) vector (Stratagene) to yield plasmid pDH205. A plasmid that contained CaCLA4 flanking sequences joined with BqlII sites was then created by PCR using the divergent oligodeoxynucleotide primers OEL109 (5'-GAAGATCTTGTAATCAATGTTCCCGTGGA-3' (SEQ ID NO:3) and OEL110 (5'-GAAGATCTCATCGTGATATTAAATCCGAT-3' (SEO NO:4); newly introduced BqlII sites are underlined) and plasmid pDH205 as template. The amplified DNA was cleaved with BqlII and ligated with a 4 kb BamHI-BqlIIfragment of a hisG-URA3-hisG cassette derived from plasmid pCUB-6 (Fonzi, W. A. & Irwin, M. Y. (1993) Genetics 134, 717-728) to yield plasmid pDH210. This

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plasmid was linearized with PstI and SacI and transformed into the Ura C. albicans strain CAI4 (Fonzi, W. A. & Irwin, M. Y. (1993) Genetics 134, 717-728) to replace the coding region of one of the chromosomal CaCLA4 alleles with the hisG-URA3-hisG cassette by recombination. Ura+ transformants homologous selected on Ura medium, and integration of the cassette into the CaCLA4 locus was verified by Southern blot analysis. Spontaneous Ura derivatives were then selected on medium containing 5-fluoroorotic acid. These clones were screened by Southern blot hybridization to identify those which had lost the URA3 gene by intrachromosomal recombination mediated by the hisG repeats. This procedure was then repeated to delete the remaining functional allele of CaCLA4.

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To reintegrate CST20 into the genome of mutant strains, the C. albicans integration plasmid pDH190 was constructed by subcloning a KpnI to PstI fragment of CST20 into pBS-cURA3 (pBluescript KS(+) into which the C. albicans URA3 gene was cloned between the NotI and XbaI sites of the polylinker). The integration plasmid was then linearized with NsiI and transformed into C. albicans to target integration into the NsiI site of the CST20A::hisG fusion gene. Integrations were selected on Ura medium and confirmed by Southern blot analysis.

The C. albicans CST20 expression plasmid pDH188 was constructed by subcloning a SacI to PstI fragment of CST20 into plasmid pVEC carrying a C. albicans autonomously replicating sequence and URA3 as selectable marker. The C. albicans plasmid pVEC-CaCLA4 was constructed by subcloning the KpnI to SacI insert of YEp 352-CaCLA4 into plasmid pVEC.

Northern blot analyses

Northern blots of total and poly $(A)^+$ RNA from C. albicans cells were performed as described (Leberer, E. et al. (1992) *EMBO J.* 11, 4815-4824). Signals were quantified by 2-D radioimaging.

Animal experiments

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Eight week-old, male CFW-l mice (Halan-Winkelmann, Paderborn, Germany) were inoculated with 1 x 10^5 or 1 x 10^6 cells by intravenous injection. Survival curves were calculated according to the Kaplan-Meier method using the PRISMTM program (GraphPad Software Inc., San Diego) and compared using the log-rank test. A P value <0.05 was considered significant.

To quantify colony-forming *C. albicans* units in kidneys, mice were sacrificed by cervical dislocation 48 hours after injection and kidneys were homogenized in 5 ml phosphate buffered saline, serially diluted and plated on YNG medium (0.67% yeast nitrogen base, 1% glucose, pH 7.0). Histological examination of kidney sections was done with periodic acid Schiff's stain.

RESULTS

Isolation and characterization of CST20

gene was cloned by functional complementation of the pheromone signaling defect of *S. cerevisiae* cells that were deleted for the *STE20* gene. The mating defect of the *STE20* deleted *S. cerevisiae* strain YEL20 was fully complemented by introduction of the centromeric plasmid pRL53 carrying full length *CST20* (mating efficiency was 81±9% in cells expressing *CST20*, compared with 85±8% in cells expressing *STE20*; n=3). Similarly, defects in growth arrest and morphological changes in response to pheromone were completely cured by transformation with the *CST20* plasmid.

As shown in Fig. 1, nitrogen deficiency-induced pseudohyphae formation, which is blocked by disruption of STE20 in diploid cells (Liu, H., Styles, C. & Fink, G. R. (1993) Science 262, 1741-1744), was restored by introduction of the CST20 plasmid. Colonies of the diploid STE20 wild type strain L5266 (4) (Fig. 1A) and the isogenic ste20/ste20 strain HLY492 (4) transformed with either the control plasmid pRS316 (Fig. 1B), the CST20 plasmid pRL53 (Fig. 1C), or the STE20 plasmid pSTE20-5 (9) (Fig. 1D) were grown on nitrogen starvation medium (2) for 5 days at 30°C. Photomicrographs were taken with a 4x objective (bar=lmm).

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As illustrated in Fig. 2, the cytokinesis defect caused by deletion of CLA4, encoding an S. cerevisiae isoform of Ste20p (Cvrckova, F. et al. (1995) Genes 15 Dev. 9, 1817-1830), was not complemented by CST20 (Fig. However, the lethality caused by deletion of both STE20 and CLA4 (Cvrckova, F. et al. (1995) Genes Dev. 9, 1817-1830), could be rescued by CST20 (Fig. 2). 20 diploid strain YEL306 heterozygous for ste20∆ cla4∆::LEU2/CLA4 was transformed with ::TRP1/STE20 plasmid pRS316 carrying either no insert, CLA4 (pRL21), CST20 (pRL53) or STE20 (pSTE20-5), and then sporulated and dissected. No viable haploid ste201 cla41 spores 25 were obtained from transformants with the plasmid without insert, but were obtained from transformants with plasmids carrying CLA4 (Fig. 2A), STE20 (Fig. 2B) or CST20 (Fig. 2C).

Cells were grown to mid-exponential phase in YPD medium at 30°C. No viable $ste20\Delta$ $cla4\Delta$ segregants were obtained in medium containing 5-fluoro-orotic acid suggesting that the plasmids were essential for viability. Neither STE20 nor CST20 were able to suppress the morphological defect of $cla4\Delta$ cells. Photomicrographs

were taken by phase contrast with a 40x objective (bar=30 μ m).

The open reading frame of CST20 is capable of encoding a protein of 1,229 amino acids with a predicted molecular weight of 133 kDa and a domain structure characteristic of the Ste20p/p65^{PAK} family of protein kinases (Fig. 3). Numerals at the left margin indicate nucleotide and amino acid positions (Fig. 3). Nucleotide 1 corresponds to the first nucleotide of the initiation codon and amino acid 1 to the first residue of the deduced protein. The putative p21 binding domain has been shadowed, and the kinase domain has been boxed.

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The catalytic domain present in the carboxyl 15 terminal half of the protein has sequence identities of 76 and 56%, respectively, with S. cerevisiae Ste20p (Leberer, E. et al. (1992) EMBO J. 11, 4815-4824) and Cla4p (Cvrckova, F. et al. (1995) Genes Dev. 9, 1817-The amino terminal, non-catalytic region con-20 tains a sequence from amino acid residues 473 to 531 with 68% identity to the p21 binding domain of Ste20p that has been shown to bind the small GTPase Cdc42p. This region contains the sequence motif ISxPxxxxHxxH thought to be important for the interaction of the p21 25 binding domain with the GTP-bound forms of Cdc42Hs and Racl (Cvrckova, F. et al. (1995) Genes Dev. 9, 1817-The remaining non-catalytic sequences are less 1830). conserved. Unique sequences not present in Ste20p and the other members of the family are found at the amino 30 terminus and between the p21 binding and catalytic domains.

A CST20 transcript of 4.9 kb in size was detected in Northern blots. This transcript was present at similar levels in yeast cells grown in YPD at

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room temperature and germ tubes induced by a temperature shift to 37°C.

Isolation and characterization of CaCLA4

A C. albicans homolog of the S. cerevisiae CLA4 gene was cloned by functional complementation of the growth defect of S. cerevisiae cells that were deleted for the STE20 and CLA4 genes.

The open reading frame of the CaCLA4 gene is capable of encoding a protein of 971 amino acids with a predicted molecular weight of 107 kDa and a domain structure characteristic of the Ste20p family of protein kinases (Fig. 7). The catalytic domain present in the carboxyl terminal half of the protein has sequence identities of 74, 63 and 64%, respectively, with S. cerevisiae Cla4p, S. cerevisiae Ste20p and an uncharacterized open reading frame present in the S. cerevisiae genome, 65% with the C. albicans Ste20p homolog Cst20p, and 61% with rat $p65^{PAK}$ (Fig. 7). The amino terminal, noncatalytic region contains a sequence from amino acid residues 69 to 180 with similarity to pleckstrin homology (PH) domains and a sequence from amino acid residues 229 to 292 with 63% identity to the Cdc42p binding domain of S. cerevisiae Cla4p that has been shown to bind the small GTPase Cdc42p (Cvrckova, F. et al. (1995) Genes Dev. 9, 1817-1830). The remaining noncatalytic sequences are less conserved.

Chromosomal deletion of CST20

Homologous recombination was used in a multistep procedure to partially delete CST20 in a URA C. albicans strain (Fig. 4A). PCR with the divergent oligode-oxynucleotides ODH68 and ODH69 was used to partially delete the coding sequence of CST20. A hisG-URA3-hisG cassette was then inserted. The deletion was confirmed by Southern blot analyses (Fig. 4B). The genomic DNA samples digested with XhoI were from following strains:

Lane #1, CAI4 (ura3/ura3 CST20/CST20); lane 2, CDH15 (ura3/ura3 CST20/cst20\Delta::hisG-URA3-hisG); lane 3, CDH18 (ura3/ura3 CST20/cst20\Delta::hisG); lane 4, CDH22 (ura3/ura3 cst20\Delta::hisG-URA3-hisG/cst20\Delta::hisG); lane 5, CDH25 (ura3/ura3 cst20\Delta::hisG/cst20\Delta::hisG). Northern blots showed that the CST20 transcript was absent in the corresponding homozygous deletion strains.

The lateral outgrowth of hyphae from colonies grown on solid "Spider" media containing mannitol or sorbitol was completely blocked by deletion of *CST20* (Fig. 5B).

Mycelial formation was drastically reduced when the media contained galactose, mannose or raffinose. The mutant strains regained the ability to form hyphae when wild type CST20 was reintroduced by transformation with the CST20 expression plasmid pDH188 or reintegrated into the genome by targeted homologous recombination (Fig. 5C). The CST20 transcript was detected in these strains by Northern blot analysis.

Mutant strains formed hyphae when colonies were grown on "Spider" media containing either glucose or N-acetyl glucosamine. Normal hyphae formation was also observed on rice agar and on agar containing Lee's medium or 10% serum. The frequency of germ-tube formation in either liquid Lee's medium, 10% serum or liquid "Spider" media containing any of the sugars tested above, were also normal. These results indicate that Cst20p is not required for hyphae formation under all conditions but are involved in the lateral formation of mycelia on some solid surfaces.

Chromosomal deletion of CaCLA4

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Homologous recombination was used in a multistep procedure to delete both alleles of *CaCLA4* in *C. albi-cans* (Fig. 8A). Fig. 8A shows the restriction endonuclease map of *CaCLA4*. The coding sequence is indicated

by the arrow. PCR with the divergent oligodeoxynucleotides OEL109 and OEL110 was used to delete the coding sequence of CaCLA4. A hisG-URA3-hisG cassette was then inserted and a two-step procedure was used to delete both alleles of CaCLA4 by homologous recombination. The 5 endonuclease restriction sites are as follows: BamHI; Bg, BglII; E, EcoRI; H, HindIII; P, PstI; S, SacI; X, XbaI. The deletions were confirmed by Southern blot analyses (Fig. 8B). Southern blot analysis 10 with a 1.1 kb CaCLA4 fragment from PstI-XbaI as a probe. The genomic DNA samples digested with EcoRI were from following strains: Lanes: 1, CAI4 (ura3/ura3 CaCLA4/CaCLA4); 2, CDH77 (ura3/ura3 CaCLA4/cacla4/ ::hisG-URA3-hisG); 3, CDH88 (ura3/ura3 CaCLA4/cacla4/ 15 ::hisG): 4, CLJ1 (ura3/ura3 cacla4A::hisG-URA3hisG/cacla4∆::hisG); and 5, CLJ5 (ura3/ura3 cacla4∆ ::hisG/cacla4A::hisG). Northern blots showed that the CaCLA4 transcript with a size of 4.1 kb was reduced to about 40% in heterozygous CaCLA4/cacla41 cells and was absent in homozygous cacla4\(\Delta\)/cacla4\(\Delta\) deletion cells 20 (Fig. 8C). The transcript was present at about wildtype levels when the CaCLA4 gene was retransformed into the homozygous deletion cells by using an autonomously replicating plasmid carrying the CaCLA4 gene (Fig. 8C). 25 Northern blot analysis of poly(A) + RNA isolated from following strains grown in the yeast form in YPD at 30°C: Lanes: 1, SC5314 (wild-type); 2, CDH88; 3, CLJ5 transformed with pVEC; 4, CLJ5 transformed with pVEC-CaCLA4. The blot was probed with fragments specific for CaCLA4 (upper panel) or CaACT1 (lower panel) and quan-30 tified by radioimaging. Numbers at the bottom of the figure depict the relative amounts of CaCLA4 transcript in relation to the amounts of CaACT1 transcript (mean values of two independent experiments).

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We found that viability of *C. albicans* cells was not affected by deleting either one or both alleles of *CaCLA4*. Mutant cells showed the same growth behavior as wild-type cells, independently whether the cells were grown under conditions favoring either the yeast or filamentous forms. However, deletion of both *CaCLA4* alleles generated defects in cellular morphology producing a heterogeneous population of aberrantly shaped cells that were frequently multibudded and multinucleated. This phenotype indicates a defect in cytokinesis resembling the phenotype of *S. cerevisiae* cells deleted for *CLA4* (Cvrckova, F. et al. (1995) *Genes Dev.* 9, 1817-1830).

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Deletion of both CaCLA4 alleles caused defects in hyphal formation in all media and under all conditions that we investigated. When morphological switching was induced in liquid media by either serum, N-acetyl glucosamine, proline, pH increase, temperature shift, or Lee's medium, wild-type cells and cells deleted for only one or both alleles of CaCLA4 produced germ tubes after about 30 minutes. In wild-type cells and cells deleted for only one allele of CaCLA4, these germ tubes elongated and grew into long hyphae after prolonged incubation. Cells deleted for both alleles of CaCLA4 failed to produce hyphae, however. Instead, these cells produced multiple short protrusions giving rise to an aberrant morphology.

On solid media containing either serum, rice agar or mannitol, the normal formation of mycelia was completely suppressed by deletion of both *CaCLA4* alleles. This phenotype was reversed by introducing the *CaCLA4* gene on a plasmid, and deletion of only one allele had no effect.

Virulence studies

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To determine the role of Cst20p for virulence, mice were injected intravenously with wild type and mutant strains and monitored for survival and for fungal invasion into kidneys. We found that the Urastrain CAI4 was not pathogenic (Figs. 6A and B). However, infection with Ura+ wild type cells resulted in rapid mortality with a rate that was dependent on the dose of injected cells (1 x 10^6 cells in Fig. 6A, and 1 \times 10⁵ cells in Fig. 6B). Survival was significantly prolonged, however, in mice infected with Ura+ cells deleted for both alleles of CST20 (cst201/cst201 :: URA3). This effect, which was reproducible and statistically significant, was observed at high (Fig. 6A) or low (Fig. 6B) doses of infection (with P values of 0.027 and 0.001, respectively) and correlated with colony-forming units per kidney (1.5 x 106 for wild type cells and 7 x 10^5 for $cst20\Delta/cst20\Delta$:: URA3 mutant cells) after 48 hours of infection with 1 x 10^6 cells. effects on virulence could be reversed by reintroducing CST20 into the strain deleted for both CST20 alleles, and were not observed in Ura+ cells deleted for only one CST20 allele. A histological examination revealed that cells deleted for both alleles of CST20, were able to form hyphae in infected kidneys (Fig. 6C).

To investigate whether CaCla4p is required for virulence, mice were injected intravenously with wild-type and mutant *C. albicans* strains and monitored for survival and for fungal invasion into kidneys. Infections with *CaCLA4* wild-type cells (strain SC5314) resulted in rapid mortality (Fig. 9). No difference in the mortality rate was observed after infection with cells deleted for only one allele of *CaCLA4* (strain CDH77). All mice survived, however, after infection with cells deleted for both alleles of *CaCLA4* (strain

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CLJ1 and CLJ5pVEC1). This effect correlated with a reduction in the amount of colony-forming units per kidney of infected animals and was reversed by transformation of the cells with a plasmid carrying the CaCLA4 gene (strain CLJ5CaCLA4) (Fig. 9). A histological examination revealed that kidneys from injected with either wild-type cells or cells deleted for one allele of CaCLA4 were heavily infected with C. albicans cells that produced hyphae densely penetrating the animal tissue (Fig. 10, left panel), whereas kidneys from mice injected with cells deleted for both CaCLA4 alleles contained small foci of aberrantly shaped cells that frequently carried multiple protrusions (Fig. 10, right panel). The morphologies of these cells were similar to those induced by serum under in vitro conditions. Thus, the function of CaCla4p is required for morphological switching of C. albicans under in vitro and in vivo conditions and for virulence.

20 Molecular cloning of the CaCDC42 and CaBEM1 genes

A C. albicans homolog of the CaCDC42 gene was cloned by functional complementation of the temperature-sensitive growth defect of S. cerevisiae cells carrying the cdc42-1ts mutation. The growth defect was fully complemented by plasmid YEp352-CaCDC42. The open reading frame of the CaCDC42 gene is capable of encoding a protein of 191 amino acids with homology to the Rho-family of small G-proteins (Fig. 11). The highest homology is found with Cdc42p from S. cerevisiae.

A C. albicans homolog of the CaBEM1 gene was cloned by functional complementation of the growth defect of S. cerevisiae cells deleted for the BEM1 gene. This defect was fully complemented by plasmid YEp352-CaBEM1 carrying the CaBEM1 gene. The open reading frame of the CaBEM1 gene is capable of encoding a

protein of 635 amino acids with a domain structure characteristic of Bemlp (Fig. 12). CaBemlp contains two conserved SH3 domains which are most homologous to the SH3 domains of Bemlp, and also has homology to Bemlp outside of the SH3 domains.

Discussion

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In S. cerevisiae, Ste20p fulfills multiple functions during mating (Leberer, E. et al. (1992) EMBO J. 4815-4824), pseudohyphae formation (Liu. Styles, C. & Fink, G. R. (1993) Science 262, 1741-1744), invasive growth (Roberts, R. L. & Fink, G. R. (1994) *Genes* 8, 2974-2985) and Dev. cytokinesis (Cvrckova, F. et al. (1995) Genes Dev. 9, 1817-1830). CST20 expression in S. cerevisiae fully complements Thus, Cst20p has the potential to these functions. fulfill similar functions in C. albicans.

The yeast-to-hyphal transition of C. albicans is a morphological change that can be triggered by a wide variety of factors. Carbohydrates, amino acids, salts, and serum have been described as inducers of germ tube formation, as have pH changes, temperature increases and starvation, but no single environmental factor could be defined as uniquely significant in stimulating the morphological switch. Hence C. albicans appears capable of responding to many divergent environmental signals. Disruption of both CPH1 alleles, which encode a homolog of the S. cerevisiae Stel2p transcription factor (Liu, H. et al. (1994) Science 266, 1723-1726), suppressed the lateral formation of mycelia from colonies grown on solid "Spider" medium, but did not block hyphal development in other media. We have shown that C. albicans mutant cells deleted for CST20 display a similar phenotype, and that the effect of these mutations on hyphal development is dependent on the carbon source in which the cells were grown.

These observations are consistent with the idea that several signaling pathways can trigger morphogenesis in *C. albicans*. Furthermore, the behavior of *C. albicans* mutant strains deleted for either *CPH1* or *CST20* indicates that these pathways might operate independently to activate hyphal development under differing environmental conditions. *C. albicans* encounters a variety of different microenvironments during the development of superficial and systemic infections. Hence, the existence of parallel morphogenetic signaling pathways might provide a distinct advantage to this pathogen.

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Our results indicate that the pathway controlled by Cst20p is not essential for virulence in a mouse model of systemic infections. It is not inconceivable that this pathway plays a role in other forms of infections, for example in the development of superficial infections of the mucosal epithelia (thrush). An as yet undefined role of Cst20p in pathogenicity outside of the Cst20 signaling pathway is suggested, however, by prolonged survival of mice infected with cst20 deleted cells. It is unlikely that this effect is caused by defects in hyphal formation since a histological examination of infected kidneys revealed that the CST20 deleted cells are not restricted in their capacity to form hyphae.

In S. cerevisiae, Cla4p plays a role in cytokinesis and shares with Ste20p an essential function for polarized growth during budding (Cvrckova, F. et al. (1995) Genes Dev. 9, 1817-1830). Cla4p binds the Rholike small G-protein Cdc42p (Cvrckova, F. et al. (1995) Genes Dev. 9, 1817-1830) which is involved in controlling cell polarity during budding and in response to pheromone. Like Ste20p and the mammalian homolog p21-activated kinase (p65 PAK), Cla4p is able to phosphory-

late and activate myosin-I, a mechanism that may contribute to the organization of the actin cytoskeleton.

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Our finding that CaCLA4 expression in S. cerevisiae completely complements the Cla4p functions suggests that CaCla4p may have similar properties in C. albicans. Thus, CaCla4p may be required for myosin-I driven polarized growth during hyphal formation in a mechanism that may involve the C. albicans homolog of Cdc42p. Our complementation assays in S. cerevisiae suggest that CaCla4p may share an essential function with Cst20p, the C. albicans homolog of Ste20p (Figs. 6A and 6B). This notion suggests, together with our findings that null mutants of CaCLA4 are completely non-pathogenic (Fig. 10) and null mutants of CST20 are reduced in virulence (Figs. 6A and 6B), that CaCla4p and Cst20p, and proteins such as CaCdc42p and CaBemlp interacting with these protein kinases, may be valid targets for the development of antifungal agents.

The present invention will be more readily understood by referring to the following examples which are given to illustrate the invention rather than to limit its scope.

EXAMPLE I

Screening test for inhibitors of CaCla4p and Cst20p

An in vitro assay containing the proteins CaCla4p and/or Cst20p will be used to test compounds inhibiting their activity to render avirulent any fungi, which may be pathogenic.

The activity of the protein will be monitored to determine if the compounds tested do inhibit their biological activity, using myelin basic protein as a substrate.

In cases were a selective inhibition of CaCla4p and Cst20p and not to $p65^{\it PAK}$ would be desired, compounds testing positive for the inhibition of both

CaCla4p and Cst20p will be tested to determine if they also inhibit the protein $p65^{PAK}$. This would be useful in cases of pathogenic fungi infection such as for C. albicans were the fungi is to be rendered avirulent without affecting the normal protein of the patient $p65^{PAK}$.

In some cases of inflammation, it would be desirable to be provided with compounds inhibiting all three proteins, namely, CaCla4p, Cst20p and p65 PAK .

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EXAMPLE II

Screening test for inhibitors of CaCla4p and CaCdc42p interactions

15 An *in vitro* assay containing the proteins CaCla4p and CaCdc42p will be used to test compounds inhibiting their interactions.

CaCla4p may be solid phase bound and CaCdc42p will be in suspension free to interact with CaCla4p. A labeled antibody specific to CaCdc42p will be added to the assay to determine the presence of CaCdc42p bound to CaCla4p. The compounds tested to inhibit the CaCdc42p-CaCla4p interactions, should when tested positive, cause only a minute quantity of CaCdc42p to bind to CaCla4p interactions.

The analogous in vitro assay will be used to test compounds that inhibit the interaction between Cst20p and CaCdc42p.

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EXAMPLE III

Screening test for inhibitors of CaCla4p and CaBemlp interactions

An in vitro assay containing the proteins 35 CaCla4p and CaBemlp will be used to test compounds inhibiting their interactions.

CaCla4p may be solid phase bound and CaBemlp will be in suspension free to interact with CaCla4p. A

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labeled antibody specific to CaBemlp will be added to the assay to determine the presence of CaBemlp bound to CaCla4p. The compounds tested to inhibit the CaBemlp-CaCla4p interactions, should when tested positive, cause only a minute quantity of CaBemlp to bind to CaCla4p interactions.

The analogous in vitro assay will be used to test compounds that inhibit the interaction between Cst20p and CaBemlp.

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EXAMPLE IV

A two-hybrid CaCdc42p and CaCla4p interaction system in a humanized S. cerevisiae strain

This screening assay is based on the assumption that the interaction of the small G-protein CaCdc42p with its cellular targets Cst20p and CaCla4p is essential for viability of C albicans cells. This essential function is reasonable to assume based on work that has been performed in S. cerevisiae (Leberer E. et al. (1997) Embo J. **16**, 83-97). The two hybrid interaction system will use green fluorescent protein fused to the GAL1 promoter as a functional read out. This reporter gene will be integrated into a S. cerevisiae strain in which the STE20 and CLA4 genes have been replaced by the human homolog p65PAK. The CaCDC42 gene will be fused to the DNA binding domain of GAL4, and the CaCLA4 gene will be fused to the activation domain of GAL4. Interaction of the two proteins will cause green fluo-Whereas inhibitors of the interaction will rescence. suppress fluorescence.

Non-specific inhibitors of the two-hybrid interaction system will be excluded by performing a parallel screen with unrelated fusion proteins known to interact. Compounds of general toxicity or inhibitors of WO 98/18927 PCT/CA97/00809 - 27 -

the human homologs will also be excluded in this system because those compounds will not allow growth of the cells and therefore reduce the fluorescent readout in both parallel screens.

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A two-hybrid yeast strain carrying the GAL4-GFP. fusion gene is constructed. This strain will be deleted for the CLA4 gene using the TRP1 marker as described (Leberer E. et al. (1997) Embo J. 16, 83-97). The STE20 gene will be replaced by the human PAK gene as described above. To replace the CDC42 gene by its 10 human homolog, an integrating plasmid will be constructed carrying the HsCDC42 gene fused to a URA3 blaster gene and CDC42 flanking sequences. After linearization, the construct will be transformed into the PAK containing two-hybrid strain, and integrants will 15 be selected on -ura medium. The URA3 gene will then be looped out on FOA medium. The various gene disruptions and gene replacements will be verified by Southern blot analyses.

20 The two-hybrid vectors carrying the CaCDC42 gene fused to the GAL4-DNA binding domain and the CaCLA4 gene fused to the transcriptional activation domain of GAL4 will be constructed by standard procedures. facilitate the interaction of the two proteins, we will 25 use site-directed mutagenesis to create a mutation in the CAAX-box domain of CaCDC42p to prevent isoprenylation and targeting of the fusion protein to the plasma membrane. We will evaluate and optimize the assay system and adapt the assay conditions to the 30 scale used in microtiter plates for automated screening of compounds.

EXAMPLE V

Detection of the presence of C. albicans using probes

The sequences of either one of the genes CaCLA4, CST20, CaCDC42 and CaBEM1 may be used to derive probes for the detection of C. albicans using PCR techniques or hybridization assays.

EXAMPLE VI

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Use of nucleotide sequences of CaCLA4, CST20, CaCDC42 and CaBEM1 to identify homologue from other fungi

The nucleotide sequences of CaCLA4, CST20, CaCDC42 and CaBEM1 may be used to identify and clone homologues from other fungi.

EXAMPLE VII

A S. cerevisiae-based screening system using CaSte20p and the pheromone signaling pathway as drug target

In this system, we will use green fluorescent protein (GFP) under transcriptional control of a pheromone inducible promoter (FUS1) as a read out. pheromone signaling pathway and thereby the reporter gene will be induced with pheromone in two different First, in a strain in which STE20 is funcstrains. tionally replaced by the CaSTE20 gene. And second, in a strain in which STE20 is functionally replaced by the mammalian homolog PAK. Compounds that block the induction of the reporter gene in the CaSTE20 strain but not in the PAK strain are expected to be specific inhibitors of the C. albicans kinase. This assay is very specific and is a positive selection of compounds that excludes the finding of compounds with inhibitory action against the mammalian homolog PAK or compounds of general toxicity.

The FUS1 gene, including its promoter, will be isolated by the polymerase chain reaction (PCR) from

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genomic DNA of *S. cerevisiae* and fused to the *GFP* gene from *Aequoria victoria* on a yeast expression plasmid. The function of the reporter gene will be analyzed after transformation of a *MAT*a yeast strain and induction with pheromone.

The STE20 gene will be replaced in a supersensitive sst1 yeast strain by the human PAK gene using homologous recombination. For this purpose, an integrating plasmid will be constructed carrying the PAK gene fused to a URA3 blaster gene and STE20 flanking sequences. The construct will be linearized and transformed into yeast, and integrants will be selected on ura medium. The URA3 gene will then be looped out on FOA medium to gain back the ura3 marker. Correct integration of the PAK gene will be confirmed by Southern blot analysis.

The humanized strain will then be transformed with the FUS1-GFP reporter gene and analyzed for a functional signaling pathway by measuring green fluorescence after induction with pheromone. The assay system will be evaluated, optimized and adapted to the scale used in microtier plates.

EXAMPLE VIII

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Fluorescence resonance energy transfer (FRET) as probe for protein-protein interactions

The engineering of different GFP mutants with altered fluorescence characteristics allows the use of fluorescence resonance energy transfer (FRET) to probe protein-protein interactions (Heim and Tsien (1996) Curr. Biol. 6, 178-182). The FRET phenomenon consists in a fluorescence transfer between a donor and a receptor fluorochrome. If excitation and emission wavelengths are compatible, the FRET is easily measurable. The main parameter of the reaction is the distance

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between donor and receptor, which must be in the range of nanometers. This is precisely the kind of values in protein-protein interactions.

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We propose to develop a novel yeast assay system which uses FRET to measure the in vivo interaction between CaCdc42p and Cacla4p. The CaCDC42 gene will be fused to a GFP mutant that acts as donor, and the CaCLA4 gene will be fused to a mutant that acts as receptor. The yeast strain used as an expression system will be humanized as described in Example VII. Inhibitors of the interaction are expected to reduce energy transfer, and this reduction can be easily measured spectroscopically. The interaction of unrelated proteins known to interact will be used as a reference to exclude non-specific inhibitors of the assay system. Compounds inhibiting the interaction of the human homologs or of general toxicity will be excluded by inhibition of growth and therefore reduced fluorescence in both screens.

20 The CaCDC42 gene will be fused to the gene encoding the GFP^{Y66H} mutant as donor, and the CaCLA4gene will be fused to the gene encoding the GFP^{S65T} mutant as receptor (Heim and Tsien (1996), Curr. Biol. **6**, 178-182). The constructs will then be transformed 25 into the humanized yeast strain described in Example VII, and the FRET phenomenon will be analyzed in yeast cultures using fluorescence spectroscopy. The conditions for the assay will be worked out and optimized. We will adapt the assay conditions to the scale used in 30 microtiter plates for automated screening.

While the invention has been described in connection with specific embodiments thereof, it will be understood that it is capable of further modifications and this application is intended to cover any variations, uses, or adaptations of the invention following, WO 98/18927 PCT/CA97/00809 - 31 -

in general, the principles of the invention and including such departures from the present disclosure as come within known or customary practice within the art to which the invention pertains and as may be applied to the essential features hereinbefore set forth, and as follows in the scope of the appended claims.

SEQUENCE LISTING

- (1) GENERAL INFORMATION
- (i) APPLICANT: National Research Council of Canada
- (ii) TITLE OF THE INVENTION: CANDIDA ALBICANS PROTEINS
 ASSOCIATED WITH VIRULENCE AND HYPHAL FORMATION AND USES
 THEREOF
- (iii) NUMBER OF SEQUENCES: 12
- (iv) CORRESPONDENCE ADDRESS:
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 - (F) ZIP: H3A 2Y3
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Diskette
 - (B) COMPUTER: IBM Compatible
 - (C) OPERATING SYSTEM: DOS
 - (D) SOFTWARE: FastSEQ for Windows Version 2.0
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
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 - (A) APPLICATION NUMBER: 60/029,458
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- (viii) ATTORNEY/AGENT INFORMATION:
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 - (C) REFERENCE/DOCKET NUMBER: 2139-10PCT
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 - (C) TELEX:
 - (2) INFORMATION FOR SEQ ID NO:1:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 29 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA

	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:	
CGGGATC	CAG ACCAACCACT CGAACTACT	29
	(2) INFORMATION FOR SEQ ID NO:2:	
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 29 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:	
CGGGATC	CGA AGGTGAACCA CCATATTTG	29
	(2) INFORMATION FOR SEQ ID NO:3:	
	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 29 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:	
GAAGATC	TTG TAATCAATGT TCCCGTGGA	29
	(2) INFORMATION FOR SEQ ID NO:4:	
	(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 29 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
GAAGATC	TCA TCGTGATATT AAATCCGAT	29
	(2) INFORMATION FOR SEQ ID NO:5:	
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 4492 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: Genomic RNA	

PCT/CA97/00809

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence (B) LOCATION: 355...4044

(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

GTCTAACGTA TATAC ATTTTATTGT TTAGT CACAATTTAA CTATT CTCACAAGGG CTAGA	CGCGTA CACCATCTT PTTATA TCCAACCAC PTGTTT GACAGCTGA AAATAA GTTTGCAAA	TACTCCACAT TACTCCACAT GACAATTACC A AAGAGATAAA A AACAAGTTTT	CTTGACAATC CTCACTTTAA ACATATTGGA TTCAATTTTT AATAGTTTTC AATTAATATT AAAAGAATCA AGTGCTATAA AAAAATAGTA ACTGCACTTT AAATAAAGCC TATC ATG Met	60 120 180 240 300 357
			TCA ATA ACA GAT CCA Ser Ile Thr Asp Pro	405
			AAC TCT GGA ACG AGG Asn Ser Gly Thr Arg 30	453
			GAA TCA ACA CCA CTA Glu Ser Thr Pro Leu 45	501
			GCT AAT ACT TCT TCA Ala Asn Thr Ser Ser 65	549
			AAA CAA TTT GAT CAA Lys Gln Phe Asp Gln 80	597
			ACT ATA GAA TCT GGA Thr Ile Glu Ser Gly 95	645
-		Gln Ser His	AAC AAC AAC AAC Asn Asn Asn Asn 110	693
			TCA AGT GAA GGC GAT Ser Ser Glu Gly Asp 125	741
			CCA GGG ACA TTC AAT Pro Gly Thr Phe Asn 145	789
			AAT GAA AAA CAG TAC Asn Glu Lys Gln Tyr 160	837
			AGT AAA GAT TCG TAT Ser Lys Asp Ser Tyr 175	885

			CCC Pro 185					933
			ATA Ile					981
			TTA Leu					1029
			CCG Pro				 	1077
			CTG Leu					1125
 			ACT Thr 265			 		1173
			CCT Pro					1221
			ATT Ile					1269
			TTG Leu					1317
	 		CAA Gln					1365
			CAA Gln 345					1413
			TCG Ser					1461
			ACC Thr					1509
			ATC Ile					1557
			ACA Thr					1605

			TCA Ser					1653
			GTC Val 440					1701
			TCA Ser					1749
			AAT Asn					1797
			GGC Gly					1845
			AGA Arg					1893
			CCA Pro 520					1941
			AAC Asn					1989
			AGT Ser					2037
			GGT Gly					2085
			AGT Ser					2133
			GAA Glu 600					2181
			AGT Ser					2229
			TAT Tyr					2277

		ATT Ile						2325
		ATG Met						2373
		GAT Asp						2421
		GTT Val 695						2469
		AAG Lys						2517
		CCT Pro						2565
		CAT His						2613
		ACA Thr						2661
		GCT Ala 775						2709
		CCA Pro						2757
		CCA Pro						2805
		ACT Thr						2853
		TAT Tyr						2901
		GAG Glu 855						2949
		AAT Asn						2997

Gln Asn Asn			AGT AAT ATT GCT Ser Asn Ile Ala 895	Pro Pro
			GGA TCT GGT GGT Gly Ser Gly Gly 910	
		Ile Ala Gln	AAG AAA CGA GAA Lys Lys Arg Glu 925	
			TTA AAG ACA ATT Leu Lys Thr Ile 940	
			TTA GTT AAA ATT Leu Val Lys Ile	
Gly Ala Ser			GAT GTT CGT GAT Asp Val Arg Asp 975	Lys Ser
			GAA CAA CAA CCT Glu Gln Gln Pro 990	
		Leu Val Met	AAA GGT AGT CTG Lys Gly Ser Leu 1005	
			TTA AAA GGT GAT Leu Lys Gly Asp 1020	
			CTT ACC GAT ATA Leu Thr Asp Ile 5	
			GTT GTA TGT CGT Val Val Cys Arc 105	Glu Thr
	Leu Lys Phe Leu		GGG GTT ATC CAT Gly Val Ile His 1070	
		Leu Asn Met	GAT GGT AAC ATC Asp Gly Asn Ile 1085	
			GAA ATC AAT CTC Glu Ile Asn Leu 1100	

			GTG Val		Thr					Ala					Ser	3717
			TAT Tyr 1125	Gly					Val					Ile		3765
			ATG Met					Pro					Glu			3813
		Ala	TTA Leu				Ala					Pro				3861
	Pro		TCT Ser			Tyr					Phe					3909
			GAC Asp		Asn					Ala					His	3957
			ATT Ile 1205	Thr					Val					Pro		4005
			Ala					Met						'GAA'I	GG TG	4056
TTCT TATT TTTTC AATA GTCT	PACTO PCTTT PTATA CTTTTT ATATT PTGAA	GCT (FGA ATT STORY) FCT (FAT ACA A	GTCAA ATTAT IGTAT GTGTA AGCTT	ATATA PTATI PTTAI AGATO PGACI PTACO	T TO TG TI TA TA TA TA	GCTA AGTO ATATA ATGTA	ATTT GTAG ATTTT AGTAF GGTGG	CCA AGA TCA TAA AGA	ATTCT ATTTT ATTT# AGTT# AGCTG	CGT TAC AGTA ACT STAA	TTCT TAGT TTTA TGTT	'ATTT 'ATAT CTT <i>I</i> 'CAAC CTTT	CT A TT T ACA C BAC A CCC C	TTTTC TTTTT TGC# GTG# TAT#	CCTAAA CGTTTT PATTCA AGTATC AATGGA AGAAAA STATGA	4116 4176 4236 4296 4356 4416 4476 4492

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1230 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Ser Ile Leu Ser Glu Asn Asn Pro Thr Pro Thr Ser Ile Thr Asp 1 5 10 15

Pro Asn Glu Ser Ser His Leu His Asn Pro Glu Leu Asn Ser Gly Thr 20 25 30

Arg	Val	Ala 35	Ser	Gly	Pro	Gly	Pro 40	Gly	Pro	Glu	Val	Glu 45	Ser	Thr	Pro
Leu	Ala 50	Pro	Pro	Thr	Glu	Val 55	Met	Asn	Thr	Thr	Ser 60	Ala	Asn	Thr	Ser
Ser 65	Leu	Ser	Leu	Gly	Ser 70	Pro	Met	His	Glu	Lys 75	Ile	Lys	Gln	Phe	Asp 80
	Asp			85		_			90	_	_			95	
_	Ser		100		_	_		105					110		
	Asn	115					120					125			_
_	Asp 130		_			135				_	140		_		
145					150					155					Gln 160
Ī	Thr			165					170	_				175	
-	Ser		180					185	_				190		
	Asn Leu	195					200					205			
	210					215					220				Lys
225	Ser				230					235					240
	His			245	_				250					255	
	Ser	_	260					265					270		
	Pro	275	_				280			_		285			
	290 Leu					295					300				
305	Leu	_			310					315					320
	Asn		_	325	_				330					335	
	Ser		340					345					350		
		355				_	360					365			Asn
	370		_			375					380				His
385					390					395			_		400 Glu
				405					410					415	Lys
			420					425					430		
_	_	435					440					445			Phe
-	450		_			455					460				Leu
465					470				_	475					Asn 480
ALA	Lys	Hls	ьeu	Ala 485	Hls	val	GТĀ	тте	Asp 490	Asp	Asn	етА	ser	Tyr 495	rnr

Gly	Leu	Pro	Ile 500	Glu	Trp	Glu	Arg	Leu 505	Leu	Ser	Ala	Ser	Gly 510	Ile	Thr
Lys	Lys	Glu 515		Gln	Gln	His	Pro 520		Ala	Val	Met	Asp 525		Val	Ala
Phe	Tyr 530	Gln	Asp	Thr	Ser	Glu 535		Pro	Asp	Asp	Ala 540		Phe	Lys	Lys
Phe 545	His	Phe	Asp	Asn	Asn 550	Lys	Ser	Ser	Ser	Ser 555	Gly	Trp	Ser	Asn	Glu 560
	Thr			565					570			_		575	
Gly	Gly	Gly	Gly 580	Ala	Pro	Ser	Ser	Pro 585	His	Arg	Thr	Pro	Pro 590	Ser	Ser
Ile	Ile	Glu 595	Lys	Asn	Asn	Val	Glu 600	Gln	Lys	Val	Ile	Thr 605	Pro	Ser	Gln
	Met 610			_		615					620				
His 625	Glu ·	Asp	Asn	Ala	Thr 630	Gln	Tyr	Thr	Pro	Arg 635	Thr	Pro	Thr	Ser	His 640
	Gln			645					650				_	655	
	Thr		660					665			_		670		
	Ser	675				_	680			_		685			
	Lys 690					695					700	Ī		_	
Ile 705	Ser	Ser	Lys	Ser	Leu 710	Lys	Ser	Met	Arg	Ser 715	Arg	Lys	Ser	Gly	Asp 720
Lys	Phe	Thr	His	Ile 725	Ala	Pro	Ala	Pro	Pro 730	Pro	Pro	Ser	Leu	Pro 735	Ser
Ile	Pro	Lys	Ser 740	Lys	Ser	His	Ser	Ala 745	Ser	Leu	Ser	Ser	Gln 750	Leu	Arg
Pro	Ala	Thr 755	Asn	Gly	Ser	Thr	Thr 760	Ala	Pro	Ile	Pro	Ala 765	Ser	Ala	Ala
Phe	Gly 770	Gly	Glu	Asn	Asn	Ala 775	Leu	Pro	Lys	Gln	Arg 780	Ile	Asn	Glu	Phe
Lys 785	Ala	His	Arg	Ala	Pro 790	Pro	Pro	Pro	Pro	Leu 795	Ala	Pro	Pro	Ala	Pro 800
Pro	Val	Pro	Pro	Ala 805	Pro	Pro	Ala	Asn	Leu 810	Leu	Ser	Glu	Gln	Thr 815	Ser
Glu	Ile	Pro	Gln 820	Gln	Arg	Thr		Pro 825		Gln	Ala	Leu	Ala 830	Asp	Val
Thr	Ala	Pro 835	Thr	Asn	Ile	Tyr	Glu 840	Ile	Gln	Gln	Thr	Lys 845	Tyr	Gln	Glu
Ala	Gln 850	Gln	Lys	Leu	Arg	Glu 855	Lys	Lys	Ala	Arg	Glu 860	Leu	Glu	Glu	Ile
Gln 865	Arg	Leu	Arg	Glu	Lys 870	Asn	Glu	Arg	Gln	Asn 875	Arg	Gln	Gln	Glu	Thr 880
Gly	Gln	Asn	Asn	Ala 885	Asp	Thr	Ala	Ser	Gly 890	Gly	Ser	Asn	Ile	Ala 895	Pro
Pro	Val	Pro	Val 900	Pro	Asn	Lys	Lys	Pro 905	Pro	Ser	Gly	Ser	Gly 910	Gly	Gly
Arg	Asp	Ala 915		Gln	Ala	Ala	Leu 920		Ala	Gln	Lys	Lys 925		Glu	Glu
Lys	Lys 930		Lys	Asn	Leu	Gln 935		Ile	Ala	Lys	Leu 940		Thr	Ile	Cys
Asn 945	Pro	Gly	Asp	Pro	Asn 950		Leu	Tyr	Val	Asp 955		Val	Lys	Ile	Gly 960

Gln	Gly	Ala	Ser	Gly 965	Gly	Val	Phe	Leu	Ala 970	His	Asp	Val	Arg	Asp 975	Lys
Ser	Asn	Ile	V al 980	Ala	Ile	Lys	Gln	Met 985	Asn	Leu	Glu	Gln	Gln 990	Pro	Lys
	Glu	995				1	1000				-	1005			
	Asn 1010)				1015	5				1020)			
Trp 1025		Ile	Met	Glu	Tyr 1030		Glu	Gly	Gly	Ser 1035		Thr	Asp	Ile	Val 1040
Thr	His	Ser	Val	Met 1045		Glu	Gly	Gln	Ile 1050		Val	Val	Cys	Arg 1055	
Thr	Leu	Lys	Gly 1060		Lys	Phe	Leu	His 1065		Lys	Gly	Val	Ile 1070		Arg
Asp	Ile	Lys 1075		Asp	Asn	Ile	Leu 1080		Asn	Met	Asp	Gly 1085		Ile	Lys
Ile	Thr 1090		Phe	Gly	Phe	Cys 1095		Gln	Ile	Asn	Glu 1100		Asn	Leu	Lys
Arg 1105		Thr	Met	Val	Gly 1110		Pro	Tyr	Trp	Met 1115		Pro	Glu	Ile	Val 1120
Ser	Arg	Lys	Glu	Tyr 1125		Pro	Lys	Val	Asp 1130		Trp	Ser	Leu	Gly 1135	
Met	Ile	Ile	Glu 1140		Leu	Glu	Gly	Glu 1145		Pro	Tyr	Leu	Asn 1150		Thr
Pro	Leu	Arg 1155		Leu	Tyr	Leu	Ile 1160		Thr	Asn	Gly	Thr 1165		Lys	Leu
Lys	Asp 1170		Glu	Ser	Leu	Ser 1175		Asp	Ile	Arg	Lys 1180		Leu	Ala	Trp
Cys 185		Gln	Val	Asp	Phe 1190		Lys	Arg	Ala	Asp 1195		Asp	Glu	Leu	Leu 1200
His	Asp	Asn	Phe	Ile 1205		Glu	Суѕ	Asp	Asp 1210		Ser	Ser	Leu	Ser 1215	
Leu	Val	Lys	Ile 1220		Arg	Leu	Lys	Lys 1225		Ser	Glu	Ser	Asp 1230)	

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3496 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 432...3344
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

GAATTCTTTT	TAGAAGAGAA	AGAAAAAATT	СССАААААА	AAAGATTTCA	TTTAATTCCA	60
CGGGAACATT	GATTACAACC	ACGTCAACAG	TTTCCCTTTT	ATATTGAAAT	CAACATTCAA	120
TTTTGTCTTT	${\bf TTTTTTTTTT}$	CATTGATTTT	TCCCCAATCT	TTTTATCTTC	ATATTAATAT	180
TGGATATCAA	TTACTAATAC	TGTCAGGGAT	AGTTTAGTAA	ATATTTACAT	TCTCCATTCA	240
ATCCTAAATT	TTTTTTTATA	TAGCTAGTTT	TTGGTTGAAA	TAAAAAAAA	AGGGGGAAGG	300
AAGTTTTTT	TTCTATTTAT	TTAATTGTTT	TGATTCCAAC	CATATTGTAT	ATTTGTCTTG	360

TCCTCAAAAG A	ATG ACA AGT	ATT TAT AC	A TCA GAT T	CTATATTA TTGAA TG AAA AAC CAT Leu Lys Asn His 10	'AGA 470
	Pro Pro Pro			T GGC TCA GGT T Gly Ser Gly	
				T AAT ATT GTT a Asn Ile Val	
				CC AAA CCT ATT Tr Lys Pro Ile 60	
Leu Asn Ile A				G GTT CAT GTT p Val His Val 75	
				A CGG TTT ATG s Arg Phe Met 90	
	Lys Thr Leu .			A CCA TAT TCT u Pro Tyr Ser	
				A TTC CCA CTA er Phe Pro Leu	
				G TAT AGC AAA y Tyr Ser Lys 140	
Ser Gln Ser P				T AAA TCA ATT in Lys Ser Ile 155	:
				G CTA GAT GCA p Leu Asp Ala 170	
	Cys Pro Leu			T AAT AGT GGT n Asn Ser Gly	
				A ACC AAT GGT u Thr Asn Gly	
				A TTA TTA TCT y Leu Leu Ser 220	

GTG Val								1142
CAT His								1190
TTA Leu 255								1238
GAG Glu								1286
TAT Tyr								1334
TCA Ser								1382
AAT Asn								1430
TGG Trp 335								1478
CGA Arg								1526
GGC Gly								1574
GGT Gly								1622
GTT Val								1670
AGA Arg 415								1718
TCT Ser								1766
CAA Gln								1814

CAA T		ln							1862
TTA TO	er L								1910
CCT A									1958
TCA A' Ser Mo 510									2006
AAT A									2054
GAT GO		ys							2102
CAG C	ln G								2150
CAG CAG Gln G.									2198
TCA G Ser V 590									2246
AAT T Asn L			 						2294
GTG C Val L		Ĺys							2342
CCT G Pro A	lla S								2390
GAA C Glu A 6			 	 	 	 		 	2438
GTG G Val V 670									2486

					GCG Ala			2534
					GAT Asp			2582
					GTT Val			2630
					TAT Tyr 745			2678
					GGT Gly			2726
					GAA Glu			2774
					CAT His			2822
					GTT Val			2870
					TGT Cys 825			2918
					ACA Thr			2966
					GAA Glu			3014
					GAA Glu			3062
					CTT Leu			3110
					TCG Ser 905			3158
					AGA Arg			3206

						TCA Ser						3254
						TTA Leu						3302
						ACA Thr 965				TAGA	AGATTG	3353
TGT	TAT	GT 1	'AGT'I	TT T	'GGA'I			 	 		STGTTG ATTTTT	3413 3473 3496

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 971 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met 1	Thr	Ser	Ile	Tyr 5	Thr	Ser	Asp	Leu	Lys 10	Asn	His	Arg	Arg	Ala 15	Pro
Pro	Pro	Pro	Asn 20	Gly	Ala	Ala	Gly	Ser 25	Gly	Ser	Gly	Ser	Gly 30	Ser	Gly
Ser	Gly	Ser 35	Gly	Ser	Gly	Ser	Leu 40	Ala	Asn	Ile	Val	Thr 45	Ser	Ser	Asn
Ser	Leu 50	Gly	Val	Thr	Ala	Asn 55	Gln	Thr	L y s	Pro	Ile 60	Gln	Leu	Asn	Ile
Asn 65	Ser	Ser	Lys	Arg	Gln 70	Ser	Gly	Trp	Val	His 75	Val	Lys	Asp	Asp	Gly 80
Ile	Phe	Thr	Ser	Phe 85	Arg	Trp	Asn	Lys	Arg 90	Phe	Met	Val	Ile	Asn 95	Asp
Lys	Thr	Leu	Asn 100	Phe	Tyr	Lys	Gln	Glu 105	Pro	Tyr	Ser	Ser	Asp 110	Gly	Asn
Ser	Asn	Ser 115	Asn	Thr	Pro	Asp	Leu 120	Ser	Phe	Pro	Leu	Tyr 125	Leu	Ile	Asn
Asn	Ile 130	Asn	Leu	Lys	Pro	Asn 135	Ser	Gly	Tyr	Ser	Lys 140	Thr	Ser	Gln	Ser
Phe 145	Glu	Ile	Val	Pro	Lys 150	Asn	Asn	Asn	Lys	ser 155	Ile	Leu	Ile	Ser	Val 160
Lys	Thr	Asn	Asn	Asp 165	Tyr	Leu	Asp	Trp	Leu 170	Asp	Ala	Phe	Thr	Thr 175	Lys
Cys	Pro	Leu	Val 180	Gln	Ile	Gly	Glu	Asn 185	Asn	Ser	Gly	Val	Ser 190	Ser	Ser
His	Pro	His 195	Leu	Gln	Ile	Gln	His 200	Leu	Thr	Asn	Gly	Ser 205	Leu	Asn	Gly
Asn	Ser 210	Ser	Ser	Ser	Pro	Thr 215	Ser	Gly	Leu	Leu	Ser 220	Ser	Ser	Val	Leu

Thr 225	Gly	Gly	Asn	Ser	Gly 230	Val	Ser	Gly	Pro	Ile 235	Asn	Phe	Thr	His	Lys 240
Val	His	Val	Gly	Phe 245	Asp	Pro	Ala	Ser	Gly 250	Asn	Phe	Thr	Gly	Leu 255	
Asp	Thr	Trp	Lys 260	Ser	Leu	Leu	Gln	His 265		Lys	Ile	Thr	Asn 270		Asp
Trp	Lys	Lys 275	Asp	Pro	Val	Ala	Val 280		Glu	Val	Leu	Glu 285		Tyr	Ser
Asp	Ile 290		Gly	Gly	Asn	Ser 295		Ala	Gly	Thr	Pro 300		Gly	Ser	Pro
Met 305		Asn	Ser	Lys	Thr 310		Asn	Asn	Asn	Asn 315		Pro	Asn	Asn	Tyr 320
	Ser	Thr	Lys	Asn 325		Val	Gln	Glu	Ala 330		Leu	Gln	Glu	Trp 335	
Lys	Pro	Pro	Ala 340	Lys	Ser	Thr	Val	Ser 345		Phe	Lys	Pro	Ser 350		Ala
Ala	Pro	Lys 355	Pro	Pro	Thr	Pro	Tyr 360		Leu	Thr	Gln	Leu 365		Gly	Ser
Ser	His 370	Gln	His	Thr	Ser	Ser 375	Ser	Gly	Ser	Leu	Pro 380		Ser	Gly	Asn
Asn 385	Asn	Asn	Asn	Asn	Ser 390	Thr	Asn	Asn	Asn	Asn 395	Thr	Lys	Asn	Val	Ser 400
Pro	Leu	Asn	Asn	Leu 405	Met	Asn	Lys	Ser	Glu 410	Leu	Ile	Pro	Ala	Arg 415	Arg
Ala	Pro	Pro	Pro 420	Pro	Thr	Ser	Gly	Thr 425	Ser	Ser	Asp	Thr	Tyr 430	Ser	Asn
_		435	Gln	_	_		440	_				445			_
Thr	Asp 450	Ser	Ser	Gln	Gln	Gln 455	Gln	Gln	Gln	Lys	Gln 460	His	Gln	Tyr	Gln
Gln 465	Lys	Ser	Gln	Gln	Gln 470	Gln	Gln	Gln	Pro	Gln 475	Gln	Pro	Leu	Ser	Leu 480
		_	Gly	485					490					495	
			Gly 500					505			Ī	_	510		
Ser	Lys	Ile 515	His	Pro	Asp	Leu	Lys 520	Ile	Gln	Gln	Gly	Thr 525	Asn	Asn	Tyr
Ile	Lys 530	Ser	Ser	Gly	Thr	Asp 535	Ala	Asn	Gln	Val	Asp 540	Gly	Asp	Ala	Lys
Gln 545	Phe	Ile	Lys		Phe 550		Leu	Gln	Leu	Lys 555	_	Ser	Gln	Gln	Gln 560
Leu	Ala	Ser	Lys	Gln 565	Pro	Ser	Pro	Pro	Ser 570	Ser	Gln	Gln	Gln	Gln 575	Gln
Lys	Pro	Met	Thr 580	Ser	His	Gly	Leu	Met 585	Gly	Thr	Ser	His	Ser 590	Val	Thr
Lys	Pro	Leu 595	Asn	Pro	Val	Asn	Asp 600	Pro	Ile	Lys	Pro	Leu 605	Asn	Leu	Lys
Ser	Ser 610	Lys	Ser	Lys	Glu	Ala 615	Leu	Asn	Glu	Thr	Leu 620	Gly	Val	Leu	Lys
Thr 625	Pro	Ser	Pro	Thr	Asp 630	Lys	Ser	Asn	Lys	Pro 635	Thr	Ala	Pro	Ala	Ser 640
Gly	Pro	Ala	Val	Thr 645	Lys	Thr	Ala	Lys	Gln 650	Leu	Lys	Lys	Glu	Arg 655	Glu
Arg	Leu	Asn	Asp 660	Leu	Gln	Ile	Ile	Ala 665	Lys	Leu	Lys	Thr	Val 670		Asn
Asn	Gln	Asp 675	Pro	Lys	Pro	Leu	Phe 680	Arg	Ile	Val	Glu	Lys 685	Ala	Gly	Gln

Gly Ala Ser Gly Asn Val Tyr Leu Ala Glu Met Ile Lys Asp Asn Asn 695 700 Arg Lys Ile Ala Ile Lys Gln Met Asp Leu Asp Ala Gln Pro Arg Lys 710 715 Glu Leu Ile Ile Asn Glu Ile Leu Val Met Lys Asp Ser Gln His Lys 730 Asn Ile Val Asn Phe Leu Asp Ser Tyr Leu Ile Gly Asp Asn Glu Leu 745 750 Trp Val Ile Met Glu Tyr Met Gln Gly Gly Ser Leu Thr Glu Ile Ile 760 765 Glu Asn Asn Asp Phe Lys Leu Asn Glu Lys Gln Ile Ala Thr Ile Cys 775 780 Phe Glu Thr Leu Lys Gly Leu Gln His Leu His Lys Lys His Ile Ile 790 795 His Arg Asp Ile Lys Ser Asp Asn Val Leu Leu Asp Ala Tyr Gly Asn 805 810 815 Val Lys Ile Thr Asp Phe Gly Phe Cys Ala Lys Leu Thr Asp Gln Arg 820 825 830 Asn Lys Arg Ala Thr Met Val Gly Thr Pro Tyr Trp Met Ala Pro Glu 835 840 845 Val Val Lys Gln Lys Glu Tyr Asp Glu Lys Val Asp Val Trp Ser Leu 855 Gly Ile Met Thr Ile Glu Met Ile Glu Gly Glu Pro Pro Tyr Leu Asn 870 875 Glu Glu Pro Leu Lys Ala Leu Tyr Leu Ile Ala Thr Asn Gly Thr Pro 885 890 Lys Leu Lys Lys Pro Glu Leu Leu Ser Asn Ser Ile Lys Lys Phe Leu 900 905 Ser Ile Cys Leu Cys Val Asp Val Arg Tyr Arg Ala Ser Thr Asp Glu 920 925 Leu Leu Glu His Ser Phe Ile Gln His Lys Ser Gly Lys Ile Glu Glu 935 940 Leu Ala Pro Leu Leu Glu Trp Lys Lys Gln Gln Lys His Gln Gln 950 955 His Lys Gln Glu Thr Leu Asp Thr Gly Phe Ala 965

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1031 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 271...843
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

CAACCAAACC	AACTTTCATC	CTTCTACCAA	TATCTTCAAC	AAAAGTTTTA	TTCAATACTA	60
	AACAGTGTTA					120
	TATAATCATA			• •		180
	CACTTTTCCT					240

GCTCATAAAT AATTAATATA TCCATATATC ATG CAA ACT ATA AAA TGT GTT GTT Met Gln Thr Ile Lys Cys Val Val 1 5	294
GTC GGT GAT GGT GCC GTT GGT AAA ACT TGC TTA TTA ATC TCG TAT ACC Val Gly Asp Gly Ala Val Gly Lys Thr Cys Leu Leu Ile Ser Tyr Thr 10 15 20	342
ACT AGT AAA TTT CCA GCT GAT TAT GTT CCT ACT GTT TTT GAT AAT TAT Thr Ser Lys Phe Pro Ala Asp Tyr Val Pro Thr Val Phe Asp Asn Tyr 25 30 35 40	390
GCT GTA ACC GTG ATG ATA GGA GAC GAA CCA TTT ACC TTG GGA TTA TTT Ala Val Thr Val Met Ile Gly Asp Glu Pro Phe Thr Leu Gly Leu Phe 45 50 55	438
GAT ACT GCT GGT CAA GAA GAT TAC GAC AGA TTA AGG CCT TTG TCA TAT Asp Thr Ala Gly Gln Glu Asp Tyr Asp Arg Leu Arg Pro Leu Ser Tyr 60 65 70	486
CCA TCG ACT GAT GTA TTC CTT GTT TGT TTT TCC GTC ATT TCT CCC GCT Pro Ser Thr Asp Val Phe Leu Val Cys Phe Ser Val Ile Ser Pro Ala 75 80 85	534
TCG TTT GAA AAT GTT AAA GAA AAA TGG TTC CCA GAA GTT CAT CAC CAT Ser Phe Glu Asn Val Lys Glu Lys Trp Phe Pro Glu Val His His His 90 95 100	582
TGT CCC GGT GTG CCA ATA ATT ATT GTC GGT ACC CAA ACT GAT TTA CGA Cys Pro Gly Val Pro Ile Ile Val Gly Thr Gln Thr Asp Leu Arg 105 110 115 120	630
AAC GAT GAT GTT ATT TTA CAG AGA TTG CAC AGA CAA AAA TTG TCC CCA Asn Asp Asp Val Ile Leu Gln Arg Leu His Arg Gln Lys Leu Ser Pro 125 130 135	678
ATC ACC CAG GAA CAG GGT GAA AAA TTG GCT AAG GAA TTG AGA GCT GTC Ile Thr Gln Glu Gln Gly Glu Lys Leu Ala Lys Glu Leu Arg Ala Val 140 145 150	726
AAG TAT GTT GAG TGT TCT GCA TTG ACT CAA AGA GGA TTG AAA ACA GTG Lys Tyr Val Glu Cys Ser Ala Leu Thr Gln Arg Gly Leu Lys Thr Val 155 160 165	774
TTT GAC GAG GCT ATA GTA GCT GCA TTA GAA CCT CCT GTA ATT AAA AAA Phe Asp Glu Ala Ile Val Ala Ala Leu Glu Pro Pro Val Ile Lys Lys 170 175 180	822
TCG AAA AAG TGT ACT ATT TTA TAGGTCGGCG ATACTAGAAG ATAGAGGATA TTGG Ser Lys Lys Cys Thr Ile Leu 185 190	877
AAATAGGGCA TACATGAGAT ATTGAATATC TATCATTAAA TATATAATTA GTTTTTTCT AAAACCTATC TTTAGGTTTG ATCTCGTTTG ATGTGTTGGG CTGTTTCGCA AAACAGTGTT CCAATCAATA AAAAGATGTG TGTAAGACTC TAGA	937 997 1031

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 191 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Met Gln Thr Ile Lys Cys Val Val Val Gly Asp Gly Ala Val Gly Lys 10 Thr Cys Leu Leu Ile Ser Tyr Thr Thr Ser Lys Phe Pro Ala Asp Tyr 20 25 Val Pro Thr Val Phe Asp Asn Tyr Ala Val Thr Val Met Ile Gly Asp 40 Glu Pro Phe Thr Leu Gly Leu Phe Asp Thr Ala Gly Gln Glu Asp Tyr 55 Asp Arg Leu Arg Pro Leu Ser Tyr Pro Ser Thr Asp Val Phe Leu Val 70 75 Cys Phe Ser Val Ile Ser Pro Ala Ser Phe Glu Asn Val Lys Glu Lys 90 Trp Phe Pro Glu Val His His Cys Pro Gly Val Pro Ile Ile Ile 105 Val Gly Thr Gln Thr Asp Leu Arg Asn Asp Asp Val Ile Leu Gln Arg 120 Leu His Arg Gln Lys Leu Ser Pro Ile Thr Gln Glu Gln Gly Glu Lys 135 Leu Ala Lys Glu Leu Arg Ala Val Lys Tyr Val Glu Cys Ser Ala Leu 150 155 Thr Gln Arg Gly Leu Lys Thr Val Phe Asp Glu Ala Ile Val Ala Ala 170 Leu Glu Pro Pro Val Ile Lys Lys Ser Lys Lys Cys Thr Ile Leu 185

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2231 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 291...2195
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

AAGCTTGTTT CTTATCTCCT TAGTATATTG TTTTACAACA CCACATACAC ATACACATAT

AGCCTTCATT AGCCTTCATT TTGACATATT TCAATAACAA TCAAGAACTA CAAGTCATAA

CTGACACACA TATAATATCT TAATTGTTAT TATAAATTTA TTCTTGATTA GATTTTAGAC

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GGGCAGAAAC AAAAACGGAA AATCCAACTC ATCCCCGATA ACTACACACA TCTATATTAA ATCATCTATT AGTCTATCAG TTATATCTCC CTCCCCTTTT CTTCTAACAA ATG ATT Met Ile 1	240 296
AAG ACG TTT CGG AAA AGT AAA AGA CTG TCG AGT AAT TCA AGT TCA CCC Lys Thr Phe Arg Lys Ser Lys Arg Leu Ser Ser Asn Ser Ser Pro 5 10 15	344
AAG AAA ACA ATA TCT CGA GTA TCA TCA ACT TCA AGT AAT CAA ACA TCT Lys Lys Thr Ile Ser Arg Val Ser Ser Thr Ser Ser Asn Gln Thr Ser 20 25 30	392
CAT GAT GGA ATA TTA CAA TCA CCT AAA AAA GTC ATT AGA GCT CTA TAT His Asp Gly Ile Leu Gln Ser Pro Lys Lys Val Ile Arg Ala Leu Tyr 35 40 45 50	440
GAT TAT GAA CCT CAA GGT CCT GGA GAA TTG AAA TTT TTC AAA GGA GAT Asp Tyr Glu Pro Gly Pro Gly Glu Leu Lys Phe Phe Lys Gly Asp 55 60 65	488
TTT TTC CAT GTA TTA AAT GAT GTT GAT GAT GAA TTA CAT AAA GAA GCG Phe Phe His Val Leu Asn Asp Val Asp Asp Glu Leu His Lys Glu Ala 70 75 80	536
GAA CGT AAT GGA TGG ATA GAA GCA ACA AAT CCA ATG ACT CAA CTT AAA Glu Arg Asn Gly Trp Ile Glu Ala Thr Asn Pro Met Thr Gln Leu Lys 85 90 95	584
GGG ATG GTC CCC ATT AGT TAT TTT GAA ATA TTT GAT CGA TCT CGT CCT Gly Met Val Pro Ile Ser Tyr Phe Glu Ile Phe Asp Arg Ser Arg Pro 100 105 110	632
ACA GTT ACA GCA TCA TCA AAC AGT TTT ACA AAT TCC ATT GAT ATT CAA Thr Val Thr Ala Ser Ser Asn Ser Phe Thr Asn Ser Ile Asp Ile Gln 115 120 125 130	680
CAT CAA CAT CAA CAA GGA ATT CAC AAT GGA ACA GGA AAT CGA AAT TTA His Gln His Gln Gln Gly Ile His Asn Gly Thr Gly Asn Arg Asn Leu 135 140 145	728
AAT CAA ACA TTA TAT GCT GTT ACA CTA TAT GAA TTT AAA GCT GAA CGA Asn Gln Thr Leu Tyr Ala Val Thr Leu Tyr Glu Phe Lys Ala Glu Arg 150 155 160	7 76
GAT GAA TTG GAT ATA ATG CCT AAT GAA AAT TTA ATT ATT TGT GCA Asp Asp Glu Leu Asp Ile Met Pro Asn Glu Asn Leu Ile Ile Cys Ala 165 170 175	824
CAT CAT GAT TAT GAA TGG TTT ATT GCC AAA CCA ATA AAT CGA TTA GGT His His Asp Tyr Glu Trp Phe Ile Ala Lys Pro Ile Asn Arg Leu Gly 180 185 190	872
GGA CCA GGT TTA GTA CCT GTT TCT TAT GTT AAA ATA ATT GAT CTT TTA Gly Pro Gly Leu Val Pro Val Ser Tyr Val Lys Ile Ile Asp Leu Leu 195 200 210	920

		TAT Tyr						968
		AAT Asn						1016
		AAA Lys						1064
		ACT Thr						1112
		ACT Thr 280						1160
		AGT Ser						1208
		ACT Thr						1256
		CAA Gln						1304
		GAA Glu						1352
		GGA Gly 360						1400
		AAA Lys						1448
		TCT Ser						1496
		TTT Phe						1544
		AGT Ser						1592
		TTT Phe 440						1640

	CAA Gln													1688
	AAT Asn													1736
	TCA Ser													1784
	ACC Thr 500													1832
	CAA Gln							_						1880
	TTA Leu													1928
	TTT Phe													1976
	CAA Gln													2024
	GAT Asp 580													2072
	CTG Leu													2120
	TTT Phe		 _	_	_	 _			_		_		_	2168
	TGT Cys						TAA	ACAG	AGA :	rcaa:	raag?	AG AG	GAGAGA	2222
GAG	AGAC	AT												2231

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 635 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Met Ile Lys Thr Phe Arg Lys Ser Lys Arg Leu Ser Ser Asn Ser Ser 10 Ser Pro Lys Lys Thr Ile Ser Arg Val Ser Ser Thr Ser Ser Asn Gln 25 Thr Ser His Asp Gly Ile Leu Gln Ser Pro Lys Lys Val Ile Arg Ala 40 Leu Tyr Asp Tyr Glu Pro Gln Gly Pro Gly Glu Leu Lys Phe Phe Lys Gly Asp Phe Phe His Val Leu Asn Asp Val Asp Asp Glu Leu His Lys Glu Ala Glu Arg Asn Gly Trp Ile Glu Ala Thr Asn Pro Met Thr Gln 90 Leu Lys Gly Met Val Pro Ile Ser Tyr Phe Glu Ile Phe Asp Arg Ser 105 Arg Pro Thr Val Thr Ala Ser Ser Asn Ser Phe Thr Asn Ser Ile Asp 120 Ile Gln His Gln His Gln Gln Gly Ile His Asn Gly Thr Gly Asn Arg 135 140 Asn Leu Asn Gln Thr Leu Tyr Ala Val Thr Leu Tyr Glu Phe Lys Ala 150 155 Glu Arg Asp Asp Glu Leu Asp Ile Met Pro Asn Glu Asn Leu Ile Ile 165 170 Cys Ala His His Asp Tyr Glu Trp Phe Ile Ala Lys Pro Ile Asn Arg 185 Leu Gly Gly Pro Gly Leu Val Pro Val Ser Tyr Val Lys Ile Ile Asp 200 Leu Leu Asn Pro Asn Ser His Tyr Thr Ser Ile Asp Thr Ser Arg Arg 215 220 Ser Gln Val Ile Gln Val Ile Asn Gly Phe Asn Ile Pro Thr Val Glu 230 235 Gln Trp Lys Asn Gln Thr Ala Lys Tyr Gln Ala Ser Thr Ile Pro Leu 250 Gly Ser Ile Ser Gly Ser Gly Thr Pro Pro Thr Ser Ala Asn Ser Gln 265 270 Tyr Phe Asp Asn His Thr Met Thr Ser Asn Arg Ser Ser Leu Gly Ser 280 285 Ser Ile Ser Ile Ile Glu Ala Ser Val Asp Ser Tyr Gln Leu Asp His 295 300 Gly Arg Tyr Gln Tyr Ser Ile Thr Ala Arg Leu Asn Asn Gly Arg Ile 310 315 Arg Tyr Leu Tyr Arg Tyr Tyr Gln Asp Phe Tyr Asp Leu Gln Val Lys 325 330 Leu Leu Glu Leu Phe Pro Tyr Glu Ala Gly Arg Ile Glu Asn Ser Lys 340 345 Arg Ile Ile Pro Ser Ile Pro Gly Pro Leu Ile Asn Val Asn Asp Ser 360 Ile Ser Lys Leu Arg Arg Glu Lys Leu Asp Tyr Tyr Leu Ser Asn Leu Ile Ala Leu Pro Ser His Ile Ser Arg Ser Glu Glu Val Leu Lys Leu 390 395 Phe Asp Val Leu Asp Asn Gly Phe Asp Arg Glu Thr Asp Ala Ile Asn 405 410

Lys Arg Phe Ser Lys Pro Ile Ser Gln Lys Ser Asn Ser His Gln Asp 425 Arg Leu Ser Gln Tyr Ser Asn Phe Asn Val Leu Gln Gln Gln Gln 440 445 Gln Gln Gln Gln Gln Tyr Ala His His Ser Arg Gly Ser Asp Asn 455 460 Ser Pro Thr Asn Glu Ser Ser Gly Ser Asn Leu Ile Asn Ser Ser Ser 470 475 His Asn Asp Ser Ser Leu Ser Ser Ser Pro Pro Pro Pro Pro Gln 485 490 495 Thr Val Thr Thr Asn Thr Thr Asn Thr Thr Ile Thr Thr Asp Ser 500 505 Ser Ser Lys Gln Pro Lys Ala Lys Val Lys Phe Tyr Phe Asp Asp 520 Ile Phe Val Leu Leu Ile Pro Thr Asn Leu Arg Leu Gln Asp Leu Lys 535 Ser Lys Leu Phe Lys Arg Leu Glu Leu Asp Ile Thr Tyr Lys Tyr Glu 550 555 Lys Pro Asp Gln Gln Gln Lys Pro Thr Ser Glu Ser Ile His Leu Phe 565 **570** Leu Lys Asn Asp Phe Glu Asp Phe Leu Ile Glu Asn Glu Thr Ser Asn 580 585 Asn Asn Asn Leu Glu Ile Asp Phe Glu Asn Glu Ile Ile Lys Glu Lys 600 Leu Gly Glu Phe Glu Val Asn Asp Asp Glu Lys Phe Gln Ser Ile Leu 615 Phe Asp Lys Cys Lys Leu Met Val Leu Val Tyr 630

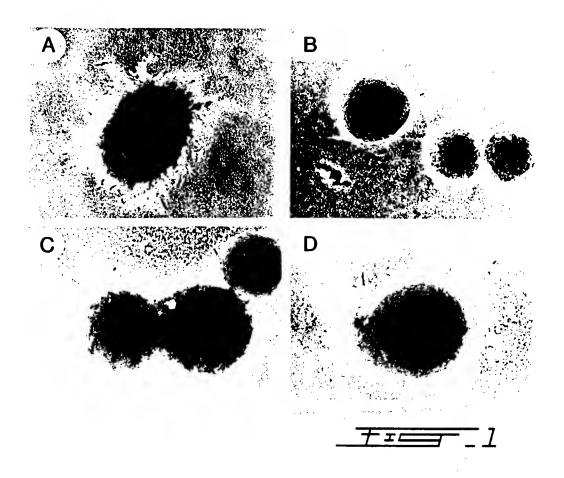
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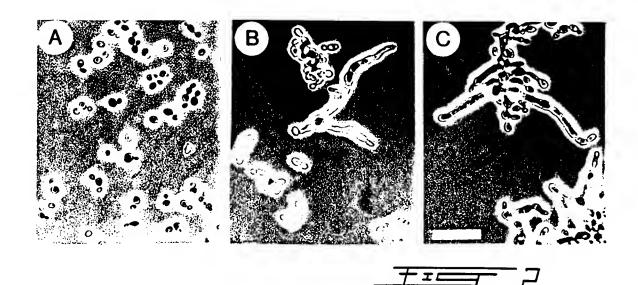
WE CLAIM:

- 1. An *in vitro* screening test for compounds to inhibit the biological activity of at least one protein selected from the group consisting of CaCla4p, Cst20p, CaCdc42p and CaBemlp, which comprises:
- a) at least one of said proteins; and
- b) means to monitor the biological activity of said at least one protein; thereby compounds are tested for their inhibiting potential.
- 2. The screening test of claim 1, wherein the inhibition of the interactions between CaCla4p and CaCdc42p is determined.
- 3. The screening test of claim 1, wherein the inhibition of the interactions between Cst20p and CaCdc42p is determined.
- 4. The screening test of claim 1, wherein the inhibition of the interactions between CaCla4p and CaBemlp is determined.
- 5. The screening test of claim 1, wherein the inhibition of the interactions between Cst20p and CaBemlp is determined.
- 6. A method for determining at least one gene involved in filamentous growth associated with virulence, which comprises using one protein selected from the group consisting of CaCla4p, Cst20, CaCdc42p and CaBemlp to determine said gene.

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CCCCGTCAACTATCACAAATACTAGTGCGACGTCAAGAAATACTTCGGGAACA PRQLSQNTSATSRNTSGT attttatttatttagtttagattatccaccactgacaattaccaattactattcaattaatattcacaatttaactatttgatttgacagctgaaaagagataaaagaatc CTGGAAACAGACACATTGAGTAGTGCCACAAATTCTCCA caacaagtgataaataccaatgcaacaaatagttcatcttcactagaaccattgggggttggcataaattcaaatctgtctcctaaaaggggaaaaagcggaaaggggaag z I × œ ı Ω × Ö (L) O I CTTAATGC CACCCC CGAGGT'R × ы Ö GCAAGATTATCTGC A R L S P TCA S ព្គម ព្គម × S Ω Ŋ , 1000 z ω z TCAAATC S N Η Ω Н Ö ш Ω ATGAGCATACTTTCAGAGAACAATCCTACACATCAATAACAGATCCAAATGAGTCTTCT(
M S I L S E N N P T P T S I T D P N E S S Ω̈́ ¤ H z ט A, Д ACATCA Ø Ω Σ Ö O Ω Д z ı × ഗ œ CGAGATGACGAAAAT. R D D E N Ω I **3GACCTGAAGTTGAATCAACACCACTAGCACCCCCAACTGAGGTCATGAATA** ACTICIGIGO I S V ເນ ы ט Ω S S AGAAA E ერ ш O STZ ည် လ Ω ы D, ы Ş d z Ω Ω S S O S ğ Ω O 241 81 601 721 841 961 321 361 1201 481 161 41 361 .354

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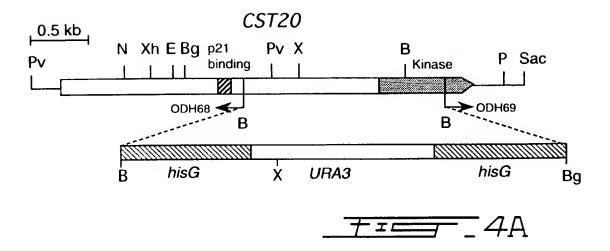
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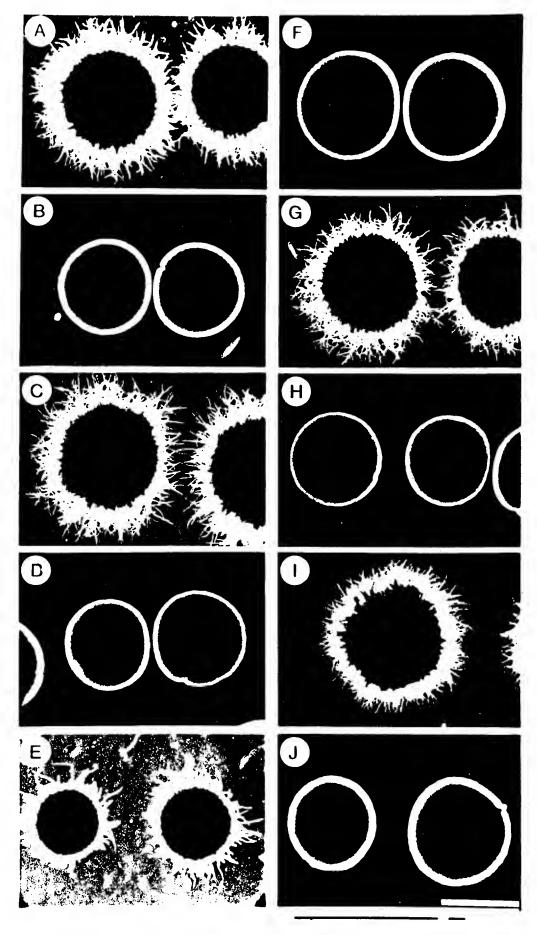
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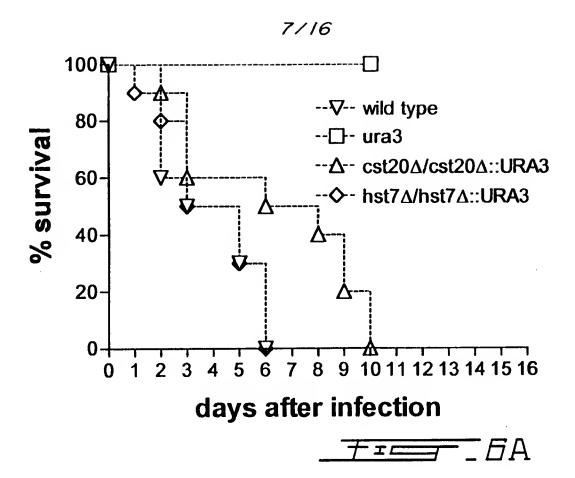
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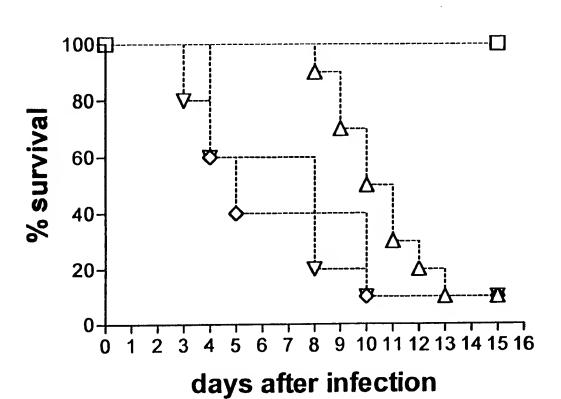
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TCTATTTATTTAATTGTTTTGATTCCAACCATATTGTATATTTGTCTTGTCAGTTATATTACTTTCTTGTTACTTAATTATTAATTGTTATTAGCTATATTGAATTGAATCCTCAAAAGA

ATGACAAGTATTTATACATCAGATTTGAAAAACCATAGACGTGCGCCACCTCCACCAAATGGGGCAGCTGGCTCTGGGTTCTGGCTCAGGTTCTAGGTTCTGGTTCTGGTTCT

GCTAATATTGTTACCAGTTCTAATAGTCTTGGCGTAACGAAATCAAACCTATTCAATTAAATATAAATTCTAGCAAACGTCAATCAGGTTCATGTTAAAGATGATGGT A N I V T S S N S L G V T A N Q T K P I Q L N I N S S K R Q S G W V H V K D D G

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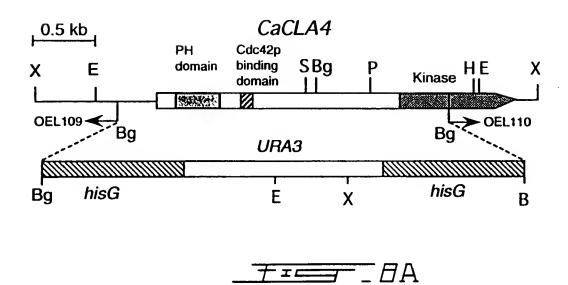
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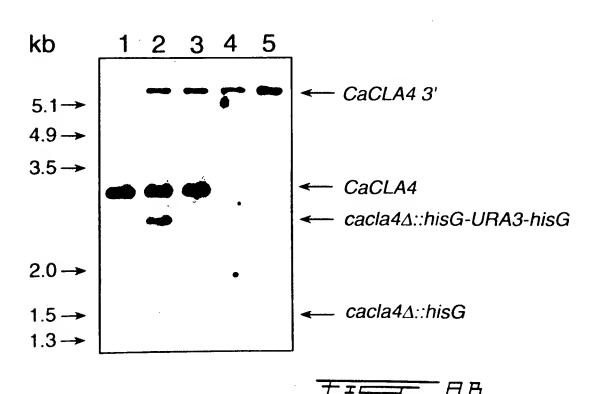
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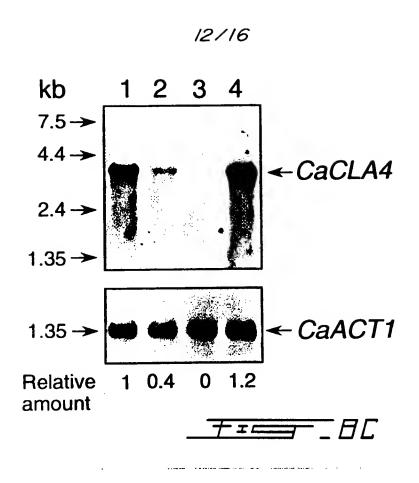


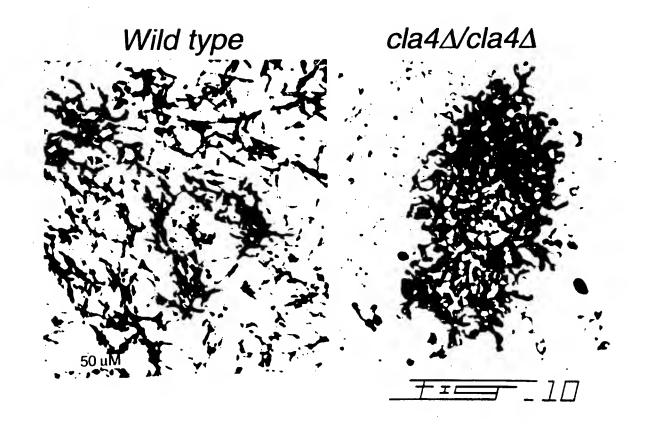
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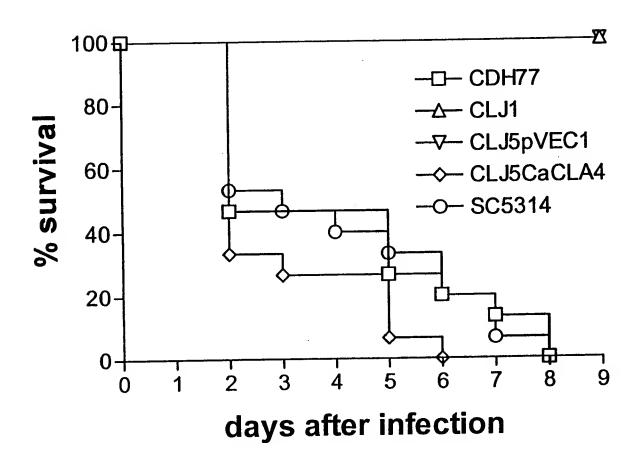


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Pathogenicity of recombinant C.albicans in mice (n=15)



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PCT/CA 97/00809 A. CLASSIFICATION OF SUBJECT MATTER IPC 6 C12N15/31 C07K C07K14/40 C12N9/12 C12N9/16 C1201/18 G01N33/68 //(C12Q1/18,C12R1:725) According to International Patent Classification (IPC) or to both national classification and IPC **B. FIELDS SEARCHED** Minimum documentation searched (classification system tollowed by classification symbols) IPC 6 CO7K C12O C12N Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Category 6 Citation of document, with indication, where appropriate, of the relevent passages Relevent to claim No. WHITE TC ET AL: "Candida albicans Α 1,6 secreted aspartyl proteinases: isoenzyme pattern is determined by cell type, and levels are determined by environmental factors." J BACTERIOL, SEP 1995, 177 (18) P5215-21, UNITED STATES, XP002057294 see abstract see page 5125, right-hand column, line 11 - line 15 see page 5220, right-hand column, paragraph 1 - paragraph 2 Χĺ Further documents are listed in the continuation of box C. Patent family members are listed in annex. Special categories of cited documents: "T" later document published after the international tiling date or priority date and not in conflict with the application but "A" document defining the general state of the art which is not cited to understand the principle or theory underlying the considered to be of particular relevance invention "E" earlier document but published on or after the international "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to document which may throw doubts on priority claim(s) or which is cited to establish the publicationdate ot another citation or other speciel reason (as specified) Involve an inventive step when the document is taken elone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such docu-"O" document referring to an oral disclosure, use, exhibition or ments, such combination being obvious to e person skilled in the art. other meens document published prior to the international tiling date but later then the priority date claimed "&" document member of the same patent family Date of the actual completion of theinternational search Date of mailing of the international search report 2 March 1998 11/03/1998 Name and mailing address of the ISA Authorized officer Europeen Patent Office, P.B. 5818 Patentiaan 2

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NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016

Inte onal Application No
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